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Total Synthesis of Siomycin A: Construction of Synthetic Segments

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Abstract: The five practical segments for the total synthesis of siomycin A, that is, the dehydropiperidine segment A (5), the pentapeptide segment B (3), the dihydroquinoline segment C (6), and the β -phenylselenoalanine dipeptide segments $D(7)$ and $E(4)$, were synthesized. Segment A (5) was constructed by the coupling of the azomethine ylide and the chiral sulfinimine, followed by the stereoselective reduction of the six-membered imine function. Segment B (3) was synthesized by the phenylselenylation of the β -lactone, stereoselective vinylzinc addition to the chiral sulfinimine, and oxazoline–thioamide conversion. Segment C (6) was

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prepared by the one-pot olefination of the tetrahydroquinoline N-oxide using triflic anhydride and triethylamine, stereoselective reduction of the methyl ketone function, and regioselective Yb- (OTf) ₃-catalyzed epoxide opening by the amino group. Segments D (7) and E (4) were synthesized by coupling of the properly protected b-phenylselenoalanines.

Introduction

Thiostrepton was isolated in 1955 from the culture broth of Streptomyces azureus by the Squibb group as an antibiotic with high inhibition activities against Gram-positive bacteria.[1] Its structure assignment had been investigated for a long time by chemical degradation studies^[2] until the complete structure was elucidated by an X-ray crystallographic analysis in 1970.[3] NMR spectral studies of thiostrepton, to-

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gether with the siomycins and thiopeptins, have been carried out in detail.^[4] In 1961, siomycin A (1) was isolated from the culture broth of Streptomyces sioyaensis by the Shionogi group.[5] In 1969, siomycin B was recognized as an artifact of siomycin A (1) , and siomycin C was isolated from the same culture broth.^[6] Siomycin D_1 was also isolated from the same culture broth in 1980 as a minor component of the siomycins.[7] The structures of the siomycins were elucidated by chemical degradation studies^[8] and NMR spectral studies^[4a-d, 7,9] by comparison with that of thiostrepton. Other structurally related antibiotics, the thiopeptins,^[10] Sch 18640 ,^[11] and Sch 40832 ,^[12] were also isolated. The characteristic structure of this thiostrepton family of peptide antibiotics is the bicyclic skeleton containing a tetrasubstituted dehydropiperidine and/or piperidine moiety, a tetrasubstituted dihydroquinoline moiety, four thiazole moieties, a thiazoline moiety, dehydroamino acid moieties, and a dihydroxyisoleucine moiety (Figure 1). Fascinated by their stunningly complex structural features, we^[13] and the Nicolaou group^[14] have aimed to synthesize these antibiotics. Recently, Nicolaou and his co-workers have succeeded in the fascinating total synthesis of thiostrepton.[15] We have also succeeded in the total synthesis of siomycin A (1) .^[16] Other efforts have focused on the syntheses of the structurally simpler thiopeptide antibiotics which have the pyridine-containing monocyclic skeleton, for example, the micrococcins,[17] promothiocin A,^[18] amythiamicin D,^[19] GE2270A,^[20,21] and GE2270T^[20]

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Thiostrepton (left): R^1 = CH₂CH₃, R^2 = CH₃, R^3 = H, R^4 = CH₃, R^5 = O, R^6 = NH₂ Siomycin A (1) (left): $R^1 = CH_3$, $R^2 = R^3 = CH_2$ (dehydroalanine), $R^4 = CH_3$, $R^5 = O$, $R^6 = NH_2$ Siomycin C (left): R^1 = CH₃, R^2 = R^3 = CH₂ (dehydroalanine), R^4 = CH₃, R^5 = O, R^6 = OMe Siomycin D₁ (left): $R^1 = CH_3$, $R^2 = R^3 = CH_2$ (dehydroalanine), $R^4 = H$, $R^5 = O$, $R^6 = NH_2$ Thiopeptin B_b (left): $R^1 = CH_3$, $R^2 = CH_3$, $R^3 = H$, $R^4 = CH_3$, $R^5 = S$, $R^6 = OH$ Thiopeptin B_a (right): $R^1 = CH_3$, $R^2 = CH_3$, $R^3 = H$, $R^4 = CH_3$, $R^5 = S$, $R^6 = OH$ Sch 18640 (right): R^1 = CH₂CH₃, R^2 = CH₃, R^3 = H, R^4 = CH₃, R^5 = S, R^6 = NH₂

Figure 1. Structures of thiostrepton antibiotics.

Figure 2. Structures of thiopeptide antibiotics.

(Figure 2). The chemical and biological properties of the thiostrepton and thiopeptide antibiotics have been recently reviewed (for example, the thiostrepton and thiopeptide antibiotics interfere with bacterial protein synthesis on the ribosome).^[22] In this and the following^[23] article, we report the construction of the five practical synthetic segments and the total synthesis of siomycin A (1).

Abstract in Japanese:

Results and Discussion

Strategy for Construction of the Bicyclic Skeleton

Aimed at the total synthesis of thiostrepton, Nicolaou and co-workers adopted the strategy of first constructing the B ring, followed by elongation of the side chain, A-ring precursor incorporation (peptide coupling), and the final Aring cyclization (lactonization; Scheme 1).^[15] Our strategy for the total synthesis of siomycin $A(1)$, on the other hand, is different from the Nicolaou one; after the A-ring (segment A-C-D (2)) construction, it is coupled to segment B (3), followed by cyclization (lactamization) of the resulting coupling product and elongation of the side-chain segment E (4) onto the cyclization product (Scheme 2). We anticipated that this cyclization–elongation step would be realized in a stepwise manner after the selective deprotection of one of the two trimethylsilylethyl (TMSE) esters, or more conveniently, in a one-pot operation by the regioselective cycliza-

ペプチド性チオストレプトン系抗生物質シオマイシン A の全合成のための 重要な5個のセグメント、すなわち、デヒドロピペリジンセグメントA、ペ ンタペプチドセグメントB、ジヒドロキノリンセグメントC、β-フェニルセ レノアラニンジペプチドセグメントDとEを合成した。

Scheme 1. Nicolaou's strategy for construction of the second ring and the side-chain elongation. Alloc=allyloxycarbonyl, $Fm = 9$ -fluorenylmethyl, TES= triethylsilyl, $TBS = tert$ -butyldimethylsilyl.

Scheme 2. Our strategy for construction of the second ring and the side-chain elongation. TMSE=trimethylsilylethyl, Teoc=2-(trimethylsilyl)ethoxycarbonyl.

tion–elongation of the dicarboxylic acid secured by the simultaneous deprotection of the two TMSE esters. Other challenging tasks include the well-timed construction of the easily racemizing thiazoline ring and the tricky dehydroamino acid units (that is, four dehydroalanine groups and one trisubstituted Z olefin adjacent to the thiazoline ring), the latter of which would be obtained by the oxidative syn elimination of five phenylseleno groups^[24] during the final stage of the total synthesis.

Retrosyntheses of Segments A-C-D (2) and A (5)

Scheme 3 shows the retrosynthetic analysis of segment A-C-D (2), which is divided into the tetrasubstituted dehydropiperidine segment $A(5)$ (strategically including the L-threonine and l-alanine residues), the tetrasubstituted dihydroquinoline segment $C(6)$ (having the *L*-valine residue), and the β -phenylselenoalanine dipeptide segment D (7). Segment A (5) would be derived from the tetrasubstituted piperidine 8 by the alanine coupling and dehydrogenation. It is expected that the latter dehydrogenation would regioselectively proceed because the C2 hydrogen atom in 8 seems to be more acidic than the C6 hydrogen atom.[25] Piperidine 8 is expected to be obtained from an equilibrium mixture of the six-membered imine derivative 9 and the five-membered imine derivative 10 by chemo- and stereoselective reduction. We anticipated that the six-membered imine 9 in this mixture would be preferentially reduced owing to steric hindrance around the imine function in the five-membered imine 10. It is also expected that the stereoselectivity of this reduction would be controlled by the stereoelectronic effect as shown in Scheme 3. To synthesize the equilibrium mix-

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Scheme 3. Retrosynthetic analyses of segments A-C-D (2) and A (5). Boc=tert-butoxycarbonyl, Bpoc=1-(4-biphenyl)-1-methylethoxycarbonyl, p-Tol= p-toluene.

ture of 9 and 10, we selected the coupling reaction between the azomethine ylide derived from dehydropyrrolidine 11 and the chiral sulfinimine 12a or its epimer 12b, followed by desulfinylation. This coupling reaction may proceed by either 1,3-dipolar cycloaddition^[26] or 1,2-addition. There is a precedent for the 1,3-dipolar cycloaddition between the azomethine ylides, derived from the N-benzylidene α -aminoesters, and the chiral sulfinimines.[27, 28] The stereochemical outcome described in the literature^[27a] matches our requirement when sulfinimine 12a is used. On the other hand, when this reaction proceeds by 1,2-addition, it is possible to control the C6 stereochemistry by employing sulfinimine 12a or 12b; however, the C5 stereochemistry is unpredictable. Dehydropyrrolidine 11 was prepared from the known pyrrolidine 13 ,^[29] and the chiral sulfinimine $12a$ (or $12b$) was prepared from aldehyde 14 and the chiral sulfinamide 15a (or $15 b$).^[30]

Synthesis of dehydropyrrolidine 11 . $[13a]$ cis-1-Boc-2,5-dicarbethoxypyrrolidine $(cis-13)^{[29]}$ (Scheme 4), prepared with modification from diethyl 2,5-dibromoadipate, was hydro-

Scheme 4. Synthesis of dehydropyrrolidine 11.

Scheme 5. Coupling between dehydropyrrolidine 11 and sulfinimine 12. DIBAL=diisobutylaluminium hydride, CMD=chemical manganese dioxide.

lyzed to the acid, which was converted into the mixed anhydride with ethyl chloroformate; into this solution, gaseous NH3 was then introduced. The resulting amide was treated with the Lawesson reagent,^[31] giving thioamide cis -16 in 33% overall yield. The treatment of cis-16 with ethyl bromopyruvate^[17d] in EtOH afforded cis-17 and trans-17 in 61 and 17% yield, respectively. The treatment of cis-17 with t BuOCl^[32] in THF followed by dehydrochlorination with triethylamine provided 11 in 95% yield. In addition, trans-1- Boc-2,5-dicarbethoxypyrrolidine $(trans-13)^{[29]}$ (Scheme 4) was also transformed into 11 by the same procedure as described in the *cis* series. From thioamide *trans*-16, obtained from trans-13 in 19% yield together with the 16% yield of cis-16, a comparable yield (each 32%) of cis-17 and trans-17 was obtained, and then *trans*-17 was converted into 11 in 76% yield.

Synthesis of chiral sulfinimines 12a and 12b, and their coupling with dehydropyrrolidine 11 ^[13a] The synthesis of the chiral sulfinimines $12a$ and $12b$ began with the known 19 , [17d] which was prepared from L-threonine (18; Scheme 5). Diisobutylaluminum hydride (DIBAL) reduction of 19 followed by oxidation with manganese dioxide^[33] afforded aldehyde 14 in 73% yield. We postulated that a mixture of $LiClO₄$ –Et₃N,^[26] which seemed to be suitable for the subsequent key coupling between dehydropyrrolidines and sulfinimines, might be used for condensation of an aldehyde with a sulfinamide. Condensation of 14 in THF with the Davis sulfinamide^[34] **15a** or **15b** in the presence of LiClO₄ and Et_3N provided sulfinimine 12a or 12b, respectively. Each solution of these sulfinimines was directly used in the subsequent coupling. First, based on the results described in the literature,[27a] dehydropyrrolidine 11 and sulfinimine 12 a were selected for the coupling partners. The coupling products turned out to be the 1,2-addition product 20 and its diastereomer^[35] in 65 and 6% yield, respectively. The following transformation of the coupling product 20 led to 28, which proved to be a diastereomer of the desired 24 and has the

opposite configurations at the C2, C5, and C6 positions (see below). Therefore, sulfinimine 12b was next chosen as the coupling partner. To the above-mentioned solution of 12b in THF was added 11 at -25° C. After one day, the addition product 21 and its diastereomer^[35] were obtained in 71 and 17% yield, respectively. The ¹H NMR signal of NHSO (δ = 5.79, $J=8.7$ Hz) in 21 supported the five-membered imine structure; however, the C5 and C6 configurations could not be determined at this stage.

Synthesis of segment A (5): $[13a,e]$ After desulfinylation of 21 with TFA in EtOH, the obtained mixture of 9 and 10 was subjected to reduction with NaBH₃CN in AcOH/EtOH to afford, as we expected, piperidine 8 in 52% yield as the sole reduction product (Scheme 6). In preference to the structurally congested primary amine at C5, the oxazolidinone amine in $\bf{8}$ was protected with Boc₂O, and the following condensation with Boc-Ala-OH (22) using 2-chloro-1,3-dimethylimidazolidium hexafluorophosphate $(CIP)^{[36]}$ and 1-hydroxy-7-azabenzotriazole (HOAt), which are useful coupling reagents for structurally congested cases, afforded piperidine 23 in 78% yield (two steps). The HMBC spectrum of 23 (from H6 to C2) supported the piperidine skeleton and the NOE experiments supported the relative configuration of the piperidine ring (Scheme 6). The absolute structure of 23 (and hence 8) was confirmed by its transformation (deprotection) to 24 and comparison of its optical rotation and ¹H and 13C NMR spectra with those of a degradation product from the natural thiopeptin B_a . [10f, 37] On the other hand, dehydropyrrolidine 20 was also transformed, through 26 and 27, into 28, which has the identical relative but opposite absolute configurations within the piperidine ring to those of 24. As we expected, dehydrogenation of 23 with $t\text{BuOCl}^{[32]}$ and triethylamine gave only dehydropiperidine 25 in 95% yield.

It is interesting to note that the configuration of the piperidine ring in thiopeptin B_a (Figure 1) was confirmed by our synthesis of 24 . According to reference [10 f], the authors

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Scheme 6. Structure determination of piperidines 8 and 26. TFA = trifluoroacetic acid, DMAP = 4-(dimethylamino)pyridine, CIP = 2-chloro-1,3-dimethylimidazolidium hexafluorophosphate, HOAt=1-hydroxy-7-azabenzotriazole.

have isolated two degradation products, IIA and its C2 epimer IIB (the compound numbers described in reference $[10f]$), from thiopeptin B_a as the piperidine residue. Compound IIA was formed from IIB as an artifact under the degradation conditions. Therefore, compound IIB was the intact piperidine residue. Unfortunately, the authors could

not analyze the coupling constant $J_{2,3}$ of compound IIB because the C2 proton signal overlapped with the C6 proton signal using the 100 MHz NMR spectrometer.^[37] Instead, judging from the coupling constant $(J_{2,3}=3.5$ and 12.0 Hz) in the ¹H NMR spectrum of IIA together with the X-ray analysis of thiostrepton,^[3] they proposed that IIA was 24 and

hence the C2 configuration of the piperidine ring in natural thiopeptin B_a was S. In contrast, according to reference [41], the authors proposed the C2 configuration of the piperidine ring in thiopeptin B_a to be R based on the coupling constant $(J_{2,3}=3.5$ and 10.0 Hz) in the ¹H NMR spectrum of thiopeptin A_{1a} (thiopeptin A_{1a} is the methyl ester of thiopeptin B_a at the terminal position R^6 depicted in Figure 1). These two assignments disagree. Therefore, we did NOE experiments for both 23 and the synthetic 24 that are shown in Scheme 6 and unambiguously determined the C2, C5, and C6 relative configurations.

The stereoselectivity observed in the 1,2-addition manner of the coupling between the azomethine ylide derived from 11 and sulfinimine 12b may be interpreted as follows (Figure 3). The metal species chelates with both the sulfinyl

Figure 3. Transition-state models for coupling between azomethine ylide and sulfinimine.

oxygen atom and the azomethine ylide nitrogen atom in the transition state (TS). It seems likely that the azomethine ylide attacks the sulfinimine opposite to the p -Tol group. In this situation, TS B and TS D suffer from an electronic repulsion between the lone pairs on the sulfur and sulfimine nitrogen atom. Thus, TS A and TS C are lower in energy than TS B and TS D. In TS C, there is a steric repulsion between the R substituent of the sulfinimine and the hydrogen atom (H3) of the azomethine ylide. As a result TS A is the most likely, giving the coupling product 21 as the major product. In the case of 12 a, the same explanation is applicable.

Since it seems apparent that the ethyl esters in 25 (Scheme 6) cannot be hydrolyzed after the construction of segment A-C-D (2), which contains the lactone function, these ethyl groups were changed to the TMSE protecting groups (Scheme 7). The treatment of 8 with trimethylsilylethanol in the presence of $Ti(iPro)_{4}^{[38]}$ gave TMSE ester 29 in 75% yield. The Boc protection (93% yield) of the oxazolidinone amine in 29 followed by condensation with Bpoc-Ala-OH $(30)^{[39]}$ using CIP,^[36] HOAt, and *iPr*₂NEt afforded 31 in 84% yield. The selective deprotection of the oxazolidinone in the presence of the TMSE esters was realized

Scheme 7. Synthesis of segment A (5).

with $Cs_2CO_3^{[40]}$ in trimethylsilylethanol (80% yield) and the successive chlorination with $tBuOC[^{32}]$, and dehydrochlorination with triethylamine and DMAP gave the dehydropiperidine segment A (5) in 76% yield.

Retrosynthesis of Segment B (3)

The retrosynthetic analysis of segment B (3) is illustrated in Scheme 8. In general, thiazolines are sensitive to epimerization;[41] therefore, it is desirable to construct the thiazoline ring in the later stage of the total synthesis. We anticipated that the β -hydroxythioamide function would be suitable as the thiazoline precursor. The β -hydroxythioamide portion would be constructed from β -hydroxyamide 32 by the Wipf oxazoline–thiazoline conversion method.[41d–f] On the other hand, the (Z)-dehydroamino acid portion would be obtained by the oxidative syn elimination of the phenylseleno group.^[24,42] β -Hydroxyamide 32 is divided into carboxylic acid 33 and the dihydroxyisoleucine derivative 34. Tripeptide 33 would be obtained from three amino acids 35–37 by consecutive condensations as well as phenylselenylation of the β -lactone part. The β -lactone function acts as not only the protecting group of the carboxylic acid group during the amide formation, but also the activating group of the hydroxy group for phenylselenylation. Two amino acids, 35

Scheme 8. Retrosynthetic analysis of segment B (3).

and 36, could be obtained from l-threonine (18). The dihydroxyisoleucine derivative 34 would be obtained from the trisubstituted olefin 38 by the stereoselective dihydroxylation. The chiral olefin 38 would be obtained from the chiral sulfinimine 39 by the stereoselective addition of an organometallic reagent.[30] If this reaction mainly affords the adduct having the undesired configuration, all we have to do is use the enantiomer of 39. Sulfinimine 39 could be easily obtained by the condensation of the known thiazole aldehyde $40^{[43]}$ and the Ellman chiral sulfinamide $41^{[44]}$ using the Cs ₂-mediated sulfinimine synthesis recently developed by our group.^[45]

Synthesis of tripeptide 33:^[13c] Tripeptide 33 was synthesized by the route shown in Scheme 9. L-Threonine (18) was treated with 2-(trimethylsilyl)ethyl 4-nitrophenyl carbonate^[46] and triethylamine followed by acetonization with acetone dimethylacetal and TsOH to afford 35 quantitatively.

 β -Lactone 36 (TsOH salt) was prepared from 18 by the Vederas method.[47] After several screenings of condensation reagents, coupling of 35 with 36 was realized using benzotriazol-1-yloxytripyrrolidinophosphonium hexafluorophosphate $(PyBOP)^{[48]}$ and iPr_2NEt in DMF to give 42 in 66% yield. The crucial step in the synthesis of 33 was the b-lactone opening by phenylselenylation. This phenylselenylation was tested using 43 (derived from 36 by the Boc protection) as the model compound. First, Shirahama's conditions^[42a] $(PhSeNa)^{[49]}$ were applied to 43, resulting in failure probably because attack of the nucleophiles at the β -position of the threonine β -lactone is disfavored in contrast to the facile ring opening at the methylene of the serine-derived β -lactones.[47] Other methods available for the nucleophilic phenylselenylation include $PhSeSiMe₃+KF₁^[50] PhSeSi Me₃+ZnI₂$ ^[51] both of which had been used for the lactoneopening reactions, and PhSeSiMe₃ and/or PhSeH used for

Scheme 9. Synthesis of tripeptide 33. Ts=p-toluenesulfonyl, PyBop=benzotriazol-1-yloxytripyrrolidinophosphonium hexafluorophosphate, DMTMM= 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium chloride, NMM=N-methylmorpholine.

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opening of the oxazoline and oxazine rings.[52] Among them, the PhSeH method[52] was the best choice in view of the easy experimental procedures (PhSeH, DMF, 80° C, 2 h), quantitatively giving 44.^[53] To the best of our knowledge, this is the first example of the opening reaction of β -substituted β -lactones using PhSeH. β -Lactone 42 was then subjected to the same reaction conditions to afford the desired 45 in 94% yield. Condensation of 45 with 37 (HCl salt) using 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methylmorpholinium chloride $(DMTMM)^{[54]}$, which is convenient for amide formation in the presence of an alcohol function, and Nmethylmorpholine (NMM) in MeOH afforded tripeptide 46 in 82% yield. Hydrolysis of 46 with aqueous NaOH gave the desired tripeptide 33, which was used in the next step without purification.

Synthesis of dihydroxyisoleucine derivative 34 : $^{[13c]}$ The synthesis of the dihydroxyisoleucine derivative 34 started with the known thiazole aldehyde 40 , $[43]$ which was prepared from ethyl diethoxyacetate (47; Scheme 10). Condensation

Scheme 10. Synthesis of dihydroxyisoleucine derivative 34. NMO= N methylmorpholine N-oxide, DABCO=1,4-diazabicyclo[2.2.2]octane, $Tf=$ trifluoromethanesulfonyl.

of 40 with the Ellman chiral sulfinamide $41^{[44]}$ in CH₂Cl₂ using our recently developed method with Cs_2CO_3 as an amine-activating and dehydrating reagent $[45]$ quantitatively produced 39. The first crucial step in the synthesis of 34 was the chemo- and stereoselective addition of the organometallic reagent to the sulfinimine group of 39 in the presence of the ethoxycarbonyl group. To the vinyllithium reagent, prepared from 1.1 equiv of (Z) -2-bromo-2-butene and 2.2 equiv of tBuLi in THF or Et₂O, was added at $-78\degree$ C a solution of 1.0 equiv of 39 in THF or Et_2O , resulting in the decomposition of 39. In contrast, transmetalation of the above vinyllithium reagent (prepared in THF) to the vinylzinc reagent by the addition of 1.1 equiv of ethereal $ZnCl₂$ was realized at 0° C; to this was added 1.0 equiv of 39 in THF at -78° C. The mixture was stirred at -40° C for 6 h, affording the desired adduct 48 in about 20% yield. Fortunately, using 5.0 equiv of the vinylzinc reagent afforded 48 in 87% yield as the sole adduct. When this addition reaction was conducted at 0° C (3 h), the ratio of 48 and its diastereomer was 3:1 and the yield of isolated 48 was reduced to 55%. The stereochemistry of 48 was confirmed in the later stage (see below). To the best of our knowledge, this is the first example of the addition of the vinylzinc reagent to the chiral sulfinimine.^[55]

The next crucial step was the dihydroxylation of the trisubstituted double bond. We expected that the sulfoxidemediated intramolecular-like dihydroxylation of olefins using $OsO₄^[56]$ was applicable to allylic sulfinamides; however, only the oxidation of the sulfinamide to the sulfonamide occurred. Sulfinamide 48 was then transformed into carbamate 38 by acid treatment followed by Boc protection in 83% yield. Dihydroxylation of 38 was conducted under a variety of conditions, including the Sharpless asymmetric dihydroxylation;[57] the best result was obtained using 0.1 equiv of $OsO₄$, 3 equiv of N-methylmorpholine N-oxide (NMO), and 0.2 equiv of 1,4-diazabicyclo[2.2.2]octane $(DABCO)^{[58]}$ in 85:15 *tBuOH–H₂O* at room temperature for 12 h, affording a 2:1 mixture of 49 and its diastereoisomer, from which the desired 49 was easily separated by silica-gel column chromatography in 56% yield. The structure determination of 49 (and hence 48) was realized by its transformation into the naturally derived degradation product, thiostreptine (50) , $[2c, 8a, 10d]$ by deprotection and comparing the optical rotation and ¹H NMR spectrum. Disilylation of 49 with triethylsilyl trifluoromethanesulfonate (TESOTf) and 2,6-lutidine afforded, concomitant with cleavage of the Boc group,^[59] 34 in 92% yield.

The high selectivity of the addition of the vinylzinc reagent to sulfinimine 39 might be rationalized by the open transition-state model $(TS E)^{[30,55b,60]}$ shown in Figure 4, in which the addition occurs from the Si face of the imine. On the basis of the Davis statement,[55b] we speculate that the existence of heteroatoms in the thiazole ester disrupts and prevents formation of the chelated transition state (TS F in

Figure 4. Transition-state models for addition of an organometallic reagent to sulfinimine.

Scheme 11. Synthesis of segment B (3). DAST = diethylaminosulfur trifluoride, TBHP = tert-butylhydroperoxide, TFE = 2,2,2-trifluoroethanol.

Figure 4, Re face attack). On the other hand, the selectivity of the dihydroxylation of 38, though it was only 2:1, might be explained by considering that the carbamate group serves to deliver the oxidant to the desired face of the double bond.^[61]

Synthesis of segment B (3) **:** [13c] Condensation of carboxylic acid 33 with the dihydroxyisoleucine derivative 34 was conducted with CIP,^[36] HOAt, and *i*Pr₂NEt in CH₂Cl₂ to give pentapeptide 32 in 83% yield (Scheme 11). Now the crucial Wipf oxazoline–thiazoline conversion method was realized as follows.^[41,62] Treatment of 32 with diethylaminosulfur trifluoride $(DAST)^{[63]}$ in CH₂Cl₂ gave oxazoline 51 in 85% yield, which was subjected to H2S in 1:1 MeOH–triethylamine to afford thioamide 52 in 90% yield. Thioamide 52 was again treated with $DAST^{[63]}$ to give thiazoline 53, which was subsequently subjected to the oxidative syn elimination using tert-butylhydroperoxide (TBHP) in 2,2,2-trifluoroethanol (TFE)–CH₂Cl₂ (1:1) to afford pentapeptide 54 in 57% yield from 52. The structure of 54 was confirmed by its ${}^{1}H$ and 13C NMR spectra, including H–H COSY, HMQC, and HMBC. At this stage, we tried hydrolysis of the ethyl ester of 54 under a variety of conditions (for example, aqueous NaOH in 2:1 EtOH-dioxane, aqueous $Ba(OH)_2$ in $MeOH^[64] Me₃SiOK$ in THF^[65]); however, but not unexpectedly, the complete epimerization occurred.^[66] Furthermore, the treatment of 54 with trimethyltin hydroxide, which could be used for hydrolysis of methyl phenylacetate, $[67]$ afforded the carboxylic acid contaminated with about 20% of the epimerization product.^[66] On the other hand, deprotection of the 2-(trimethylsilyl)ethoxycarbonyl (Teoc) group of 54 with $ZnCl₂^[68]$ in nitromethane at 50 °C resulted in approximately 20% epimerization.[66] Therefore, we considered that carboxylic acid 3 or amine 55 would be the more suitable intermediate for elaboration of the pentapeptide portion usable for the total synthesis of the thiostrepton family of peptide antibiotics. To this end, thioamide 52 was treated with aqueous NaOH in 2:1 EtOH–dioxane to afford segment B (3) quantitatively. Moreover, the treatment of thioamide 52 with $ZnCl₂^[68]$ in nitromethane afforded amine 55 in 58% yield.

Retrosynthesis of Segment C (6)

It has been proposed by Floss and co-workers^[69] that the 4-(1-hydroxyethyl)quinoline-2-carboxylic acid was an intermediate in the biosynthesis of thiostrepton. We were interested in their chemical synthesis from quinoline-2-carboxylic acid.[69] This synthetic process was applied to the early stage of our synthesis of segment $C(6)$, but with slight modifications.

The retrosynthetic analysis of segment $C(6)$ is shown in Scheme 12. The regioselective opening of epoxide 56 with the l-valine derivative 57, like the biosynthetic pathway of the quinaldic acid portion,^[69] would afford segment C (6) . Epoxide 56 would be obtained by the chemo- and diastereoselective reduction of methyl ketone 58, which would be de-

Scheme 12. Retrosynthetic analysis of segment C (6).

rived from 5,6-dihydroquinoline 59 by the asymmetric epoxidation and bromination. The 7,8-double bond in 59 would be constructed through the Boekelheide rearrangement from the 5,6,7,8-tetrahydroquinoline derivative 60, of which the acetyl group would be introduced by the radical heteroaromatic substitution reaction of 61. The modified Reissert– Henze reaction of the 5,6,7,8-tetrahydroquinoline (62) gave 61.

Construction of the hydroxyethyl portion: the first route:^[13d] We have previously synthesized the dihydroquinoline segment C (66) of siomycin D₁, with the hydroxymethyl group instead of the hydroxyethyl group (Scheme 13).^[13b] Aldehyde 63 derived from 5,6,7,8-tetrahydroquinoline (62) was transformed into alcohol 64, which was subjected to the regioselective epoxide-opening reaction^[70] with the L-valine derivative 65 in the presence of $LiClO₄^[71]$ to afford 66. The intermediate aldehyde 63 (91% ee) was the starting substance for our first route. The diastereoselective methylation of aldehyde 63 was examined under a variety of conditions and the relevant results are shown in Table 1. In the case of the methylation using MeMgBr, toluene was a better solvent than ether (Table 1, entries 1 and 2). As an additive, hexamethylphosphoramide (HMPA) was better than 1,3-dimeth $v1-3,4,5,6-tetrahedron-2(1H)$ -pyrimidinone (DMPU) and N,N,N',N'-tetramethylethylenediamine (TMEDA; Table 1,

Scheme 13. Previous synthesis of segment C (66) of siomycin D_1 . Bn= benzyl.

Table 1. Diastereoselective addition of methyl group to aldehyde 63.^[a]

63	$-78 °C$ 0.5h MeO	N 5 Ή \mathbf{A} Br НC 67	H., MeO	í.,H Ή Ēг Ή HO 68
Entry	Reagent	Additive	Solvent	Ratio ^[b] of 67/68/63
1	MeMgBr		Et ₂ O	30:14:56
2	MeMgBr		toluene	56:28:16
3	MeMgBr	HMPA	toluene	68:18:14
4	MeMgBr	DMPU	toluene	21:14:65
5	MeMgBr	TMEDA	toluene	$\lfloor c \rfloor$
6	MeLi		THF	41:13:46
7	MeLi	HMPA	THF	35:8:57

[a] 1.1 equiv of methylating reagent was added at -78° C to a solution of 1.0 equiv of 63 in solvent in the presence or absence of 1.1 equiv of additive. [b] The ratio was based on ${}^{1}H NMR$ analysis of the crude products. [c] Multispot on TLC.

entries 3–5). MeLi was less effective than MeMgBr (Table 1, entries 2, 3, 6, and 7). The best conditions to introduce the methyl group to aldehyde 63 on a 100-mg scale were as follows (Table 1, entry 3): 1.1 equiv of 3_M MeMgBr in Et₂O was added at -78 °C to a solution of 1.0 equiv 63 and 1.1 equiv HMPA in toluene. After 0.5 h at -78° C, the desired addition product 67 was obtained in 48% yield together with the undesired diastereomer 68 (15%) and the recovered 63 (20%). The configuration of the newly formed chiral center in 67 was determined by the transformation of 67 into the quinoline derivative 69 by dehydrobromination with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), aromatization under acidic conditions, and methyl ether formation with $CH₂N₂$ (Scheme 14). The obtained 69 was identical to the degradation product derived from thiostrepton^[2c] and siomycin $A^{[8a]}$. The configuration at the C5 position in 63

Scheme 14. Structure determination of 67. DBU=1,8-diazabicyclo-[5.4.0]undec-7-ene.

plays a key role in the diastereoselectivity of this reaction, the details of which will be discussed later. Although the hydroxyethyl-bearing dihydroquinoline substructure was secured at this stage, the synthetic route to 67 from 62 was lengthy and the total yield was not satisfactory. Therefore, we investigated a new route including the diastereoselective reduction of a methyl ketone function.

Construction of the hydroxyethyl portion: the second route including a new olefination at the C7–C8 position of 5,6,7,8-tetrahydroquinoline: [13d] Methyl ester 61^[13b] was acetylated under the radical heteroaromatic substitution conditions^[69b] to afford 60 in 84% yield (Scheme 15), which was

Scheme 15. Synthesis of 59. MCPBA=3-chloroperoxybenzoic acid, TFAA=trifluoroacetic anhydride.

oxidized to N-oxide 70 with 3-chloroperoxybenzoic acid (MCPBA) in 88% yield. According to our reported procedure,^[13b] N-oxide **70** was next subjected to the Boekelheide rearrangement^[72] with trifluoroacetic anhydride (TFAA) followed by hydrolysis of the resulting trifluoroacetate 71 with sodium methoxide in one pot to afford 72 in 52% yield. In our previous synthesis of the siomycin D_1 segment C (66) , ^[13b] the olefin function at the C7–C8 position was introduced in a moderate yield by treatment of the alcohol corresponding to 72 with the Burgess reagent. We expected that the C7–C8 double bond could be directly obtained from 71 under nonprotic conditions. It was found that the one-pot

treatment of 71 with three equivalents of DBU at room temperature for 20 min afforded the elimination product 59, albeit in low yield (22%). Furthermore, the following three experimental results have been reported (Scheme 16):

Scheme 16. Precedent reactions of pyridine or pyridine N-oxide with sulfonic acid derivatives.

1) Matsumura reported that 2-picoline N-oxide was transformed with tosyl chloride into 2-chloromethylpyridine via 2-tosyloxymethylpyridine;^[73] 2) picoline *N*-oxides reacted with Tf₂O at -20 ^oC to give the stable N-sulfonyloxy triflate salts;^[74] and 3) 2,6-dimethylpyridine reacted with Tf₂O to afford the compound in which a methyl hydrogen atom was replaced by a [(trifluoromethyl)sulfinyl]oxy group.[75] Encouraged by these results, we anticipated that using Tf₂O instead of TFAA in the Matsumura–Boekelheide rearrangement would more efficiently afford the elimination product 59. The relevant experimental data along this line are shown in Table 2. The treatment of 70 in CH_2Cl_2 with Tf_2O and the consecutive addition of 2,6-lutidine or iPr_2NEt expectedly afforded the elimination product 59 (Table 2, entries 1 and 2). Interestingly, but unexpectedly, deoxygenation of Noxide 70 to 60 accompanied this reaction. Fortunately, we found that dilution of iPr_2NEt in CH_2Cl_2 raised the ratio of 59/60 from 3.8:1 to 5.6:1 (Table 2, entries 2 and 3). Triethylamine was found to be a more suitable base; the ratio was improved to 10:1 (Table 2, entry 4). Finally, the best result was obtained by the slow addition of a 0.45 M CH₂Cl₂ solution of triethylamine to a solution of 70 and T_f , O in CH_2Cl_2 , producing only 59 in 98% yield (Table 2, entry 6). To the best of our knowledge, this is the first example of using Tf₂O in the Matsumura–Boekelheide rearrangement.

Although the reaction mechanism for the accompanied deoxygenation remains to be solved,^[76] that for the olefination using Tf₂O and a base by the Matsumura–Boekelheide rearrangement seems to be probable, as depicted in Scheme 17. In the case of the Boekelheide rearrangement using TFAA, $^{[72]}$ the first step is the trifluoroacetylation of Noxide 70 to give 73. The trifluoroacetate anion abstracts the proton to give the unstable intermediate 74, which undergoes rearrangement to give 71; then the basic hydrolysis of 71 finally affords alcohol 72. For the Tf_2O case, N-oxide 70 is sulfonylated to give 75, which is a stable trifluoromethane-

[a] To a solution of 70 (1.0 equiv) in CH₂Cl₂ was added Tf₂O (1.2 equiv) at 0° C, then a base (5.0 equiv) was added at 0° C over time 1, and then the mixture was stirred at room temperature for time 2. [b]Yield of isolated product $(59+60)$ after silica-gel column chromatography. [c] The ratio was based on ¹HNMR analysis of the isolated products. [d] Less than one minute.

sulfonyloxy salt.^[74] The purposely added base abstracts the proton to give the unstable intermediate 76, which would undergo rearrangement to give 77 ; finally, β -elimination of 77 affords olefin 59.

Next, we turned our attention to an asymmetric epoxidation. Olefin 59 was treated with Jacobsen's reagent $78^{[13b, 77]}$ in the presence of sodium hypochlorite as the oxidant, resulting in decomposition. In this reaction, by using iodosobenzene instead of sodium hypochlorite, epoxide 79 was obtained in 54% yield together with an 18% yield of the quinoline derivative 80 (Scheme 18). The enantiomeric excess of 79 was determined to be 75% by chiral HPLC analysis. In contrast, using Katsuki's reagent $81^{[78]}$ raised both the vield of isolated 79 and its enantiomeric excess to 73% and 82%, respectively, without the formation of 80. The absolute configuration of 79 was determined in the next stage. Bromination of 79 with N-bromosuccinimide (NBS) in CCl₄ afforded 58 in 67% yield together with an 11% yield of its diastereomer 82. The absolute configuration of 58 (and hence 79) was confirmed by an X-ray crystallographic analysis (Scheme 18).^[79] The crucial selective reduction of the methyl ketone function in 58 was conducted under a variety of conditions and the results are compiled in Table 3. DIBAL and $LiBH(sBu)$ ₃ preferentially reduced the ester function (Table 3, entries 1 and 2) to give 83 and 84. Treatment of 58 with NaBH₄ in MeOH at -78° C for 19 h afforded a 96:4 mixture of the desired reduction product 67 and its stereoisomer 68, from which 67 was isolated in 95% yield (Table 3, entry 3). Other reducing reagents, $BH₃·THF$, 9-BBN, $Zn(BH_4)_2$, and $Me_4NHB(OAc)_3$, provided no better selectivity and yield (Table 3, entries 4–7). The fact that the compound 67 derived from aldehyde 63 (Table 1) was identical to that derived from methyl ketone 58 confirmed the structure of 63, which had not previously been determined.[13b]

The diastereoselectivity observed in the addition reactions to aldehyde 63 and methyl ketone 58 may be interpreted as follows (Figure 5). Aldehyde 63, coordinated with the metal species under the stated reaction conditions, seems to prefer the conformation depicted as A rather than conformation B because of steric crowding. The attack of a methyl anion seems to occur from the Si face of the aldehyde plane to avoid the bromine atom, affording the major isomer 67. In contrast, the carbonyl and pyridine planes of methyl ketone 58 seem to be twisted to avoid the steric repulsion found in conformations C and D . Among the two conformations E and F, the former would be preferable to the latter from the viewpoint of the dipole–dipole interaction. This argument is supported by an X-ray crystallographic analysis of 58 (Scheme 18). The hydride attack seems to occur from the Re face of the carbonyl plane, affording the major isomer 67.

Scheme 17. Mechanisms for the Boekelheide rearrangement and for one-pot olefination via the Matsumura-Boekelheide rearrangement.

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Scheme 18. Synthesis and structure determination of 58. NBS = N -bromosuccinimide, AIBN = 2,2'-azobisisobutyronitrile.

Table 3. Diastereoselective Reduction of 58.

tion. After investigating a variety of reaction conditions that will be precisely discussed in the accompanying article, $^{[23]}$ it was found that $Yb(OTf)$ ₃ was more suitable for this coupling reaction; 1.0 equiv of dihydroquinoline 56 (75% ee) was coupled with 2.0 equiv of L-valine 9-fluorenylmethyl (Fm) ester $57^{[82]}$ in the presence of a catalytic amount (0.2 equiv) of Yb- (OTf) ₃ in 1:2 CH₂Cl₂-H₂O at room temperature to give 87 in 48% yield together with a 7% yield of the diastereomer of 87 arising from the enantiomer of 56, a 6% yield of the regioisomer of 87, and a 13% yield of the recovered 56. After silylation of 87 with TBSOTf (96% yield), the tert-butyl ester was deprotected with B-bromocate-

6 $Zn(BH_4)_2(1.5)$ Et₂O 0 22 42:17:--:--:41 36
7 Me.NHB(OAc), (1.7) MeCN RT 20 71:6:----23 49 $Me_4NHB(OAc)_3$ (1.7) MeCN RT 20 71:6:-:-:23 49

 $9-BBN^{[c]} (2.0)$ THF -20 to RT 3 $-$ no reaction

Comparing these two routes aimed at the preparation of 67, the first one consists of 15 steps from 62 to 67 in 1.8% overall yield, and the second one consists of 10 steps in 14% overall yield. Therefore, through the second route, we could obtain sufficient amounts of the hydroxyethyl-bearing dihydroquinoline substructure.

Synthesis of segment C (6):^[13d] After silylation of 67 with tert-butyldimethylsilyl trifluoromethanesulfonate (TBSOTf) and 2,6-lutidine (82% yield), the resulting silyl ether was subjected to dehydrobromination with DBU to afford 85 in 95% yield (Scheme 19). Methyl ester 85 was treated with potassium trimethylsilanolate^[80] to afford carboxylic acid 86, which was re-esterified with Boc₂O^[81] to afford **56** in 86% yield. In the synthesis of the siomycin D_1 segment C (66; Scheme 13), we used $LiClO₄$ for the epoxide-opening reaccholborane^[83] to give segment C (6) in 79% yield.

Syntheses of Segments D (7) and E (4) ^[13e, f]

The preparation of the β -phenylselenoalanine dipeptide segments $D(7)$ and $E(4)$, the masked precursors to the labile dehydroalanine portions, started with the known Boc-lserine β -lactone 88^[84] (Scheme 20). Phenylselenylation^[42] of 88 using the procedure reported by us^[13c] (PhSeH, DMF, RT, 2 h) gave 89 , ^[15c] which was treated with 9-fluorenylmethanol[85] and 1,3-dicyclohexylcarbodiimide (DCC) in the presence of a catalytic amount of DMAP to give Fm ester 90 in 82% yield from 88. TFA treatment of 90 followed by condensation with 89 using CIP,^[36] HOAt, and $iPr₂NEt$ in CH₂Cl₂ afforded 91 in 88% yield. Treatment of 91 with 3_M

[[]a] The ratio was based on ¹H NMR analysis of the crude products. [b] Yield of isolated 67 after silica-gel column chromatography. [c] 9-BBN = 9-borabicyclo[3.3.1]nonane.

Conformation of 63 coordinated with metal species

Figure 5. Plausible explanation for diastereoselectivity in addition reactions to 63 and 58.

Scheme 19. Synthesis of segment C (6) . TMS = trimethylsilyl.

Scheme 20. Syntheses of segments D (7) and E (4). $DCC=1.3$ -dicyclohexylcarbodiimide.

HCl/AcOEt provided segment D (7), which was used in the next step without purification.

On the other hand, 89 was converted into the mixed anhydride with $CICO₂Et$, which was treated with aqueous ammonia to afford the amide. Deprotection of the Boc group with TFA followed by condensation with 89 using CIP,^[36] HOAt, and $iPr_{2}NEt$ in CH₂Cl₂ afforded 92, which was again treated with TFA to give the side-chain segment E $(4)^{[15c]}$ in 68% overall yield from 88.

Conclusions

We have synthesized the five practical segments for the total synthesis of siomycin A, that is, the tetrasubstituted dehydropiperidine segment A (5), the pentapeptide segment B (3), the tetrasubstituted dihydroquinoline segment C (6), and the β -phenylselenoalanine dipeptide segments D (7) and E (4) . Segment A (5) was constructed by the coupling of the azomethine ylide derived from 11 and the chiral sulfinimine 12b, followed by the stereoselective reduction of the six-membered imine function. Synthesis of piperidine 24 confirmed the configuration of the piperidine ring in thiopeptin B_a . Segment B (3) was synthesized by phenylselenylation of β -lactone 42, stereoselective vinylzinc addition to the chiral sulfinimine 39, and oxazoline–thioamide conversion method $(32 \rightarrow 51 \rightarrow 52$. Segment C (6) was prepared by the one-pot olefination of 70 by Matsumura–Boekelheide rearrangement using Tf_2O and triethylamine to give 59, stereoselective reduction of the methyl ketone function in 58 controlled by the stereocenter of the peri position, and regioselective epoxide opening of 56 by amine 57 in the presence of $Yb(OTf)$ ₃. Segments D (7) and E (4) were synthesized by the coupling of the properly protected β -phenylselenoalanines. With all the important synthetic segments in hand, we set about the total synthesis of siomycin A, which will be reported in the following article.[23]

Experimental Section

General

The melting points were determined on a micro-hot-stage Yanaco MP-S3 and were uncorrected. Optical rotations were measured on a JASCO DIP-360 polarimeter. IR spectra were recorded on a JASCO FT IR-200 spectrometer. ¹H and ¹³C NMR spectra were measured on a JEOL GSX-270 spectrometer, a JEOL LAMBDA 300 spectrometer, or a Varian MERCURY plus 300 spectrometer. Chemical shifts of ¹H NMR spectra are expressed in ppm relative to TMS (0 ppm) in CDCl₃ or to the solvent residual signal in CDCl₃ (7.26 ppm), CD₃OD (3.31 ppm), (CD₃)₂SO (2.50 ppm) , (CD_3) , $CO(2.05 \text{ ppm})$, $CD_3CN(1.94 \text{ ppm})$, or 4:1 $CDCl_3$ -CD3OD (7.38 ppm) as an internal standard unless otherwise noted. Chemical shifts of 13C NMR spectra are expressed in ppm relative to solvent signal in CDCl₃ (77.00 ppm), CD₃OD (49.00 ppm), $(CD₃)₂SO$ (206.26 ppm), CD₃CN (118.26 ppm), or $[D_8]THF$ (24.55 ppm) as an internal standard unless otherwise noted. Low- and high-resolution mass spectra were recorded on a JEOL GCmate (EI and FAB), JEOL Accu TOF JMS-T100 LCS (ESI), or Bruker Ultraflex (MALDI). Silica-gel TLC and preparative TLC (PTLC) were performed on a Merck 60F-254. Silica-gel column chromatography was performed on a Fuji-Davison PSQ100B. Air- and/or moisture-sensitive reactions were carried out under an atmosphere of argon with oven-dried glassware. In general, the organic solvents were purified and dried by appropriate procedures, and evaporation and concentration were carried out under reduced pressure below 30° C.

Thioamide cis-16: To a solution of cis-13 (5.00 g, 15.9 mmol) in MeOH (40 mL), 1.4-dioxane (40 mL), and H₂O (80 mL) at 0^oC was added dropwise 1m aqueous NaOH (47.7 mL). The reaction mixture was stirred at room temperature for 2 h. AcOEt (200 mL) and 1m aqueous HCl (60 mL) were added to the solution and the mixture was extracted with AcOEt (200 mL \times 3). The combined extracts were evaporated. The residue was once again dissolved in AcOEt and the mixture was filtered through celite. The filtrate was evaporated to afford the dicarboxylic acid (3.57 g) as colorless solids. IR (CHCl₃): $\tilde{\nu} = 1760, 1705, 1045, 1390 \text{ cm}^{-1}$; ¹H NMR (CDCl₃): δ = 10.05 (brs, 2H, CO₂H), 4.59 (m, 1H, pyrrolidine H-2), 4.43 (m, 1H, pyrrolidine H-5), 2.55–1.95 (m, 4H, pyrrolidine H-3, H-4), 1.45 ppm (s, 9H, Boc); ¹³C NMR (CDCl₃): δ = 177.3, 175.5, 153.6, 82.9, 60.8, 59.8, 29.1, 28.8, 28.0 ppm; HRMS (EI): m/z [M-H₂O]⁺ calcd for $C_{11}H_{15}NO_5$: 241.0950; found: 241.0932. To a solution of this dicarboxylic acid (6.70 g, 25.8 mmol) and NEt₃ (8.64 mL, 62.0 mmol) in dry THF (260 mL) at 0° C under Ar atmosphere was slowly added ClCO₂Et (5.44 mL, 56.9 mmol). The solution was stirred at room temperature for 1 h. To the reaction mixture was introduced dry $NH₃$ gas for 10 min. Then the solvent was evaporated and the residue was chromatographed on silica gel (30–50% EtOH/hexane) to afford the dicarboxamide (3.77 g) as solids. These solids contain ammonium chloride but can be used for the next reaction without further purification. ¹H NMR (CD₃OD): δ = 4.45–4.16 (m, 2H, pyrrolidine H-2, H-5), 2.46–2.19 (m, 2H, pyrrolidine H-3, H-4), 2.13–1.90 (m, 2H, pyrrolidine H-3, H-4), 1.44 ppm (s, 9H, Boc). To a solution of this dicarboxamide (3.77 g) in dry 1,4-dioxane (70 mL) under Ar atmosphere was added Lawesson's reagent (6.52 g, 16.1 mmol). The mixture was stirred at 90° C for 1.5 h. The solvent was evaporated and then saturated aqueous $NaHCO₃$ (100 mL) was added to the residue. The mixture was extracted with AcOEt (150 mL \times 3). The

combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (50–80% AcOEt/hexane) to afford cis-16 (2.84 g, 33% from cis-13) as solids: R_f 0.54 (90% AcOEt/hexane); m.p. 114–115°C; IR (KBr): $\tilde{v} = 3300$, 3130, 1685, 1630, 1440, 1390, 1365, 1245, 1160 cm⁻¹; ¹H NMR ((CD₃)₂CO): δ = 9.94 (brs, 1H, NH₂), 9.70 (brs, 1H, NH₂), 9.03 (brs, 2H, NH₂), 4.75–4.60 (m, 2H, pyrrolidine H-2, H-5), 2.46–2.25 (m, 2H, pyrrolidine H-3, H-4), 2.20–1.98 (m, 2H, pyrrolidine H-3, H-4), 1.39 ppm (s, 9H, Boc); ¹³C NMR ((CD₃)₂CO): δ = 210.9, 209.9, 154.5, 81.0, 69.4, 33.5, 33.1, 28.2 ppm; HRMS (EI): m/z [M]⁺ calcd for C₁₁H₁₉N₃O₂S₂: 289.0919; found: 289.0926.

Thioamides trans-16 and cis-16: To a solution of trans-13 (7.31 g, 23.2 mmol) in MeOH (60 mL), 1,4-dioxane (80 mL), and $H₂O$ (120 mL) at 0° C was added dropwise 1 M aqueous NaOH (69.6 mL, 69.6 mmol). The reaction mixture was stirred at room temperature for 3 h. AcOEt (50 mL) and 1m aqueous HCl (100 mL) were added to the solution and the mixture was extracted with AcOEt (200 mL \times 3). The combined extracts were evaporated. The residue was once again dissolved in AcOEt and the mixture was filtered through celite. The filtrate was evaporated to afford the dicarboxylic acid $(5.87 g)$ as colorless solids. IR $(CHCl₃)$: $\tilde{v} = 1725, 1700, 1420, 1045 \text{ cm}^{-1}$; ¹H NMR (CD₃OD): $\delta = 4.46-4.32 \text{ (m, m)}$ 2H, pyrrolidine H-2, H-5), 2.42–2.22 (m, 2H, pyrrolidine H-3, H-4), 2.13– 1.95 (m, 2H, pyrrolidine H-3, H-4), 1.43 ppm (s, 9H, Boc); 13C NMR (CD₃OD): δ = 176.2, 175.9, 155.6, 82.0, 61.0, 60.6, 30.0, 29.2, 28.5 ppm; HRMS (EI): m/z [M]⁺ calcd for C₁₁H₁₇NO₆: 259.1056; found: 259.1077. To a solution of this dicarboxylic acid (5.87 g, 22.6 mmol) and NEt_3 (7.56 mL, 54.2 mmol) in dry THF (226 mL) at 0° C under Ar atmosphere was slowly added $CICO₂Et$ (4.75 mL, 49.7 mmol). The solution was stirred at room temperature for 1.5 h. To the reaction mixture was introduced dry NH_3 gas for 10 min. The solvent was evaporated and the residue was chromatographed on silica gel (50% EtOH/hexane) to afford the dicarboxamide (4.26 g) as solids. These solids contain ammonium chloride but can be used for the next reaction without further purification. ¹H NMR (CD₃OD): δ = 4.48–4.32 (m, 2H, pyrrolidine H-2, H-5), 2.43–2.23 (m, 2H, pyrrolidine H-3, H-4), 2.26–1.84 (m, 2H, pyrrolidine H-3, H-4), 1.44 ppm (s, 9H, Boc). To a solution of this dicarboxamide $(4.26 g)$ in dry 1,4-dioxane $(83 mL)$ was added Lawesson's reagent (7.37 g, 18.2 mmol) under Ar atmosphere. The mixture was stirred at 90°C for 1.5 h. The solvent was evaporated and then saturated aqueous $NaHCO₃$ (100 mL) was added to the residue. The mixture was extracted with AcOEt (200 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (60–70% AcOEt/hexane) to afford cis-16 (1.22 g, 16% from trans-13) as solids and trans-16 (1.04 g, 19% from trans-13) as solids. trans-16: $R_f = 0.43$ (90% AcOEt/hexane); m.p. 257-258 °C; IR (KBr): $\tilde{v} = 3370, 3320, 3200, 3150, 1705, 1675, 1640, 1620, 1430, 1380,$ 1365, 1255, 1160 cm⁻¹; ¹H NMR ((CD₃)₂CO): δ = 8.88–8.52 (brm, 4H, $NH₂ \times 2$), 4.96–4.86 (m, 2H, pyrrolidine H-2, H-5), 2.72–2.50 (m, 2H, pyrrolidine H-3, H-4), 1.98–1.81 (m, 2H, pyrrolidine H-3, H-4), 1.38 ppm (s, 9H, Boc); ¹³C NMR (CD₃COCD₃): δ = 211.4, 210.6, 154.3, 80.3, 69.0, 68.7, 32.7, 31.8, 28.5 ppm; HRMS (EI): m/z [*M*]⁺ calcd for C₁₁H₁₉N₃O₂S₂: 289.0919; found: 289.0919.

Pyrrolidines cis-17 and trans-17 from cis-16: To a solution of cis-16 (1.22 g, 4.22 mmol) in dry EtOH (42 mL) in Ar atmosphere was added 85% ethyl bromopyruvate (1.25 mL, 8.44 mmol). The mixture was heated at reflux for 2 h. The solvent was evaporated and then saturated aqueous NaHCO₃ (50 mL) was added to the residue. The mixture was extracted with AcOEt (50 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (5–10% acetone/hexane) to afford cis-17 (983 mg, 61%) as solids and *trans*-17 (268 mg, 17%) as solids. *cis*-17: $R_f = 0.26$ (10% acetone/CHCl₃); m.p. 112–113 °C; IR (CHCl₃): $\tilde{v} = 1720, 1490,$ 1320, 1290, 1100 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.12 (s, 2H, thiazole H-5), 4.90 (m, 2H, pyrrolidine H-2, H-5), 4.43 (q, 4H, J=7.1 Hz, $CO_2CH_2CH_3 \times 2$), 3.46 (brs, 1H, NH), 2.58–2.38 (m, 2H, pyrrolidine H-3, H-4), 2.16–1.98 (m, 2H, pyrrolidine H-3, H-4), 1.41 ppm (t, 6H, J= 7.1 Hz, CO₂CH₂CH₃×2); ¹³C NMR (CDCl₃): $\delta = 179.3, 161.5, 147.5,$ 127.3, 61.3, 59.9, 34.1, 14.3 ppm; HRMS (EI): m/z [M]⁺ calcd for $C_{16}H_{19}N_3O_4S_2$: 381.0817; found: 381.0801; elemental analysis (%) calcd

for $C_{16}H_{19}N_3O_4S_2$: C 50.38, H 5.02, N 11.02, S 16.81; found: C 50.24, H 4.99, N 11.03, S 16.89. trans-17: $R_f = 0.47$ (10% acetone/CHCl₃); m.p. 149–150°C; IR (CHCl₃): $\tilde{v} = 1720, 1485, 1375, 1320, 1100$ cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.11$ (s, 2H, thiazole H-5), 4.96–4.88 (m, 2H, pyrrolidine H-2, H-5), 4.42 (q, 4H, $J=7.2$ Hz, $CO_2CH_2CH_3\times 2$), 3.06 (brs, 1H, NH), 2.56–2.37 (m, 2H, pyrrolidine H-3, H-4), 2.22–2.04 (m, 2H, pyrrolidine H-3, H-4), 1.41 ppm (t, 6H, $J=7.2$ Hz, $CO_2CH_2CH_3\times 2$); ¹³C NMR (CDCl₃): δ = 177.5, 161.4, 147.2, 127.4, 61.4, 59.5, 33.1, 14.3 ppm; HRMS (EI): m/z [M]⁺ calcd for C₁₆H₁₉N₃O₄S₂: 381.0817; found: 381.0844, elemental analysis (%) calcd for $C_{16}H_{19}N_3O_4S_2$: C 50.38, H 5.02, N 11.02, S 16.81; found: C 50.65, H 5.28, N 10.80, S 16.76.

Pyrrolidines trans-17 and cis-17 from trans-16: To a solution of trans-16 (1.04 g, 3.60 mmol) in dry EtOH (36 mL) in Ar atmosphere was added 85% ethyl bromopyruvate (1.06 mL, 7.20 mmol). The mixture was heated at reflux for 2 h. The solvent was evaporated and then saturated aqueous NaHCO_3 (50 mL) was added to the residue. The mixture was extracted with AcOEt (50 mL \times 3). The combined extracts were dried over Na2SO4, filtered through celite, and evaporated. The residue was chromatographed on silica gel (5–10% acetone/hexane) to afford trans-17 (441 mg, 32%) as solids and cis-17 (443 mg, 32%) as solids.

Dehydropyrrolidine 11 from cis-17: To a solution of cis-17 (741 mg, 1.94 mmol) in dry THF (20 mL) at -78° C under Ar atmosphere was added dropwise tert-butyl hypochlorite (0.231 mL, 2.04 mmol). The solution was stirred for 20 min and then NEt₃ (1.08 mL, 7.77 mmol) was added. The temperature was raised to room temperature and the reaction mixture was stirred for 4 h. The reaction was quenched with saturated aqueous $Na_2S_2O_3$ (1 mL) and saturated aqueous NaHCO₃ (30 mL). The mixture was extracted with AcOEt (50 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (60% AcOEt/hexane) to afford 11 (701 mg, 95%) as colorless solids: m.p. 144-146°C; IR (CHCl₃): \tilde{v} = 1725, 1045 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.33 (s, 1H, thiazole H-5), 8.14 (s, 1H, thiazole H-5), 5.72 (dd, J=7.5, 7.5 Hz, 1H, pyrrolidine H-5), 4.46 (q, $J=7.1$ Hz, 2H, CO₂CH₂CH₃), 4.44 (q, $J=7.1$ Hz, 2H, CO₂CH₂CH₃), 3.55–3.37 (m, 1H, pyrrolidine H-3), 3.37–3.19 (m, 1H, pyrrolidine H-3), 2.95–2.75 (m, 1H, pyrrolidine H-4), 2.44–2.23 (m, 1H, pyrrolidine H-4), 1.43 (t, $J=7.1$ Hz, 3H, $CO_2CH_2CH_3$), 1.42 ppm (t, $J=7.1$ Hz, 3H, $CO_2CH_2CH_3$); ¹³C NMR (CDCl₃): $\delta = 173.3, 171.8, 163.4, 161.3, 160.9,$ 148.4, 147.2, 130.7, 127.4, 74.4, 61.6, 61.4, 36.0, 30.4, 14.3, 14.2 ppm; HRMS (EI): m/z [M]⁺ calcd for C₁₆H₁₇N₃O₄S₂: 379.0660; found: 379.0661; elemental analysis (%) calcd for $C_{16}H_{17}N_3O_4S_2$: C 50.65, H 4.52, N 11.07, S 16.90; found: C 50.63, H 4.34, N 11.10, S 16.84.

Dehydropyrrolidine 11 from trans-17: To a solution of trans-17 (426 mg, 1.12 mmol) in dry THF (11 mL) at -78° C under Ar atmosphere was added tert-butyl hypochlorite (0.132 mL, 1.17 mmol). The solution was stirred for 20 min and then NEt_3 (0.624 mL, 4.48 mmol) was added. The temperature was raised to room temperature and the reaction mixture was stirred for 4 h. The reaction was quenched with saturated aqueous $Na₂S₂O₃$ (1 mL) and saturated aqueous NaHCO₃ (10 mL). The mixture was extracted with AcOEt $(20 \text{ mL} \times 3)$. The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (60% AcOEt/hexane) to afford 11 (321 mg, 76%) as colorless solids.

Thiazole aldehyde 14: To a solution of 19 (800 mg, 3.12 mmol) in dry toluene (21 mL) at -78 °C under Ar atmosphere was added DIBAL (1.0 M in toluene, 10.0 mL, 10.0 mmol). The reaction mixture was stirred at room temperature for 1.5 h. The reaction was quenched with H_2O (50 mL), followed by addition of pottasium sodium tartrate tetrahydrate (8.81 g, 31.2 mmol). The mixture was stirred until the emulsion disappeared. The mixture was extracted with AcOEt (50 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (50–70% acetone/hexane) to afford the alcohol (578 mg, 86%) as a syrup $\lbrack a \rbrack_D^{28}$ -22.0 (c 1.00, CHCl₃); IR (neat): $\tilde{v} = 3280, 1750, 1385, 1300, 1235, 1205,$ 1110, 1060, 980 cm⁻¹; ¹H NMR (CDCl₃): δ = 7.29 (brs, 1H, NH), 7.23 (s, 1H, thiazole H-5), 4.83 (dd, 1H, J=1.2, 6.3 Hz, oxazolidine H-4), 4.70 (brs, 2H, CH₂OH), 4.62 (dq, 1H, J=6.3, 6.3 Hz, oxazolidine H-5), 3.87 (brs, 1H, OH), 1.57 ppm (d, 1H, $J=6.3$ Hz, oxazolidine 5-Me); ¹³C NMR

(CDCl₃): δ = 170.4, 158.8, 157.1, 116.0, 80.0, 61.0, 60.3, 19.8 ppm; HRMS (EI): m/z [M]⁺ calcd for C₈H₁₀N₂O₃S: 214.0412; found: 214.0411. This alcohol (402 mg, 1.88 mmol) and CMD (2.45 g) in AcOEt (9 mL) and CH_2Cl_2 (9 mL) were stirred at room temperature for 20 h. The suspension was filtered through celite and the filter cake was washed with AcOEt. The filtrate and washings were concentrated. The residue was chromatographed on silica gel (50% acetone/hexane) to afford 14 $(340 \text{ mg}, 85\%)$ as a pale yellow syrup: $[\alpha]_{D}^{27}$ -54.0 (c 1.00, CHCl₃); IR (neat): $\tilde{v} = 1750, 1695, 1490, 1390, 1300, 1230, 1130, 1110, 1065 \text{ cm}^{-1}$; ¹H NMR (CDCl₃): $\delta = 9.96$ (s, 1H, CHO), 8.25 (s, 1H, thiazole H-5), 7.48 (br s, 1 H, NH), 4.99 (dd, $J=1.1$, 6.1 Hz, 1 H, oxazolidinone H-4), 4.69 (dt, $J=6.1, 6.1$ Hz, 1H, oxazolidinone H-5), 1.64 ppm (d, $J=6.1$ Hz, 3H, oxazolidinone 5-Me); ¹³C NMR (CDCl₃): δ = 184.1, 172.2, 158.8, 155.3, 129.2, 79.8, 60.9, 19.9 ppm; HRMS (EI): m/z [M]⁺ calcd for C₈H₈N₂O₃S: 212.0256; found: 212.0254.

Adduct 21: A solution of 14 (336 mg, 1.58 mmol), 15b (245 mg, 1.58 mmol), LiClO₄ (1.40 g, 13.2 mmol), and NEt₃ (1.84 mL, 13.2 mmol) in dry THF (13 mL) was stirred at room temperature for 5 h under Ar atmosphere to form 12b. The temperature was cooled to -40° C and then 11 (500 mg, 1.32 mmol) was added to the above solution. The mixture was stirred at -25° C for 1 day. The reaction mixture was quenched with $H₂O$ (20 mL) and extracted with AcOEt (20 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (85–100% AcOEt/ hexane) to afford 21 (678 mg, 71%) as colorless solids and its diastereomer (166 mg, 17%) as colorless solids. **21**: m.p. 110–111 °C; $[\alpha]_D^{30}$ –134.8 $(c 1.00, CHCl₃)$; IR $(CHCl₃)$: $\tilde{v} = 1765, 1730, 1480, 1420, 1340, 1100,$ 1050 cm^{-1} ; ¹H NMR (CDCl₃): $\delta = 8.32$ (s, 1H, thiazole H-5), 7.95 (s, 1H, thiazole H-5), 7.45 (d, $J=8.4$ Hz, 2H, toluene), 7.28 (d, $J=8.4$ Hz, 2H, toluene), 6.79 (s, 1H, thiazole H-5), 6.70 (d, $J=1.2$ Hz, 1H, oxazolidinone NH), 5.79 (d, J=8.7 Hz, 1H, NHSO), 4.94 (d, J=8.7 Hz, 1H, CHNHSO), 4.77 (dd, $J=1.2$, 6.6 Hz, 1H, oxazolidinone H-4), 4.51 (dq, $J=6.6$, 6.6 Hz, 1H, oxazolidinone H-5), 4.43 (q, $J=7.2$ Hz, 2H, $CO_2CH_2CH_3$), 4.39 (q, J = 7.2 Hz, 2H, CO₂CH₂CH₃), 3.22–3.06 (m, 2H, pyrrolidine H-3 \times 2), 2.66–2.46 (m, 2H, pyrrolidine H-4 \times 2), 2.43 (s, 3H, toluene), 1.62 (d, $J=6.6$ Hz, 3H, oxazolidinone 5-Me), 1.42 (t, $J=7.2$ Hz, 3H, CO₂CH₂CH₃), 1.39 ppm (t, J = 7.2 Hz, 3H, CO₂CH₂CH₃); ¹³C NMR (CDCl₃): δ = 174.4, 172.0, 168.7, 163.0, 161.2, 160.8, 158.2, 154.7, 148.5, 146.7, 141.6, 140.6, 130.9, 129.6, 128.0, 125.5, 116.5, 86.2, 79.7, 61.7, 61.4, 61.2, 59.3, 36.8, 31.8, 21.3, 19.7, 14.2 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for $C_{31}H_{33}N_6O_7S_4$: 729.1293; found: 729.1302. Diastereomer of 21: ¹H NMR (CDCl₃): $\delta = 8.35$ (s, 1H, thiazole H-5), 8.09 (s, 1H, thiazole H-5), 7.17–7.08 (m, 4H, toluene), 7.07 (s, 1H, thiazole H-5), 6.31 (br s, 1H, oxazolidinone NH), 5.97 (d, $J=10.4$ Hz, 1H, NHSO), 4.75 (d, $J=$ 10.4 Hz, 1H, CHNHSO), 4.60 (dd, J=6.0, 1.4 Hz, 1H oxazolidinone H-4), 4.56–4.37 (m, 4H, CO₂CH₂CH₃×2), 4.32 (dq, $J=6.0$, 6.0 Hz, 1H, oxazolidinone H-5), 3.09–2.95 (m, 1H, pyrrolidine H-3), 2.62–2.48 (m, 1H, pyrrolidine H-3), 2.46-2.14 (m, 2H, pyrrolidine H-4 \times 2), 2.39 (s, 3H, toluene), 1.44 (t, $J=7.0$ Hz, 3H, CO₂CH₂CH₃), 1.40 (t, $J=7.0$ Hz, 3H, $CO₂CH₂CH₃$), 1.22 ppm (d, $J=6.0$ Hz, 3H, oxazolidinone 5-Me).

Aminopiperidine 8: To a solution of 21 (500 mg, 6.86×10^{-1} mmol) in dry EtOH (7 mL) at 0°C under Ar atmosphere was added TFA (0.264 mL, 3.43 mmol). The solution was stirred at room temperature for 1 h. The reaction was quenched with aqueous NaHCO₃ (5 mL) at 0° C. The mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over Na2SO4, filtered through celite, and evaporated to afford a mixture of 9 and 10. This mixture and AcOH (0.393 mL, 6.86 mmol) were dissolved in dry EtOH (4 mL). To the mixture at 0° C was added NaBH₃CN (129 mg, 2.06 mmol) in dry EtOH (3 mL). The reaction mixture was stirred at room temperature for 2 h. The reaction was quenched with aqueous NaHCO₃ (7 mL) at 0 °C. The mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (90–100% AcOEt/hexane) to afford 8 (213 mg, 52%) as colorless solids: m.p. 86–87°C; $[a]_{D}^{30}$ +94.3 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu}$ = 1760, 1720, 1480, 1390, 1365, 1340, 1320, 1300, 1100, 1050, 1025 cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.14$ (s, 1H, thiazole H-5), 7.95 (s, 1H, thiazole H-5), 6.98 (s, 1H, thiazole H-5), 6.92 (br s, 1H, oxazolidinone NH), 4.85 (s, 1H, piperidine H-6), 4.76 (dd, J=1.2, 6.3 Hz, 1H, oxazolidinone H-4),

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4.52–4.34 (m, 2H, piperidine H-2, oxazolidinone H-5), 4.40 (q, $J = 7.2$ Hz, 2H, CO₂CH₂CH₃), 4.39 (q, $J=7.2$ Hz, 2H, CO₂CH₂CH₃), 2.78–2.40 (m, 1H, piperidine H-4b), 2.38–2.22 (m, 1H, piperidine H-3), 2.19–1.96 (m, 2H, piperidine H-4α, H-3), 1.53 (d, $J = 6.3$ Hz, 3H, oxazolidinone 5-Me), 1.39 ppm (t, J=7.2 Hz, 6H, CO₂CH₂CH₃×2); ¹³C NMR (CDCl₃): δ = 181.6, 173.7, 169.3, 161.6, 161.3, 158.3, 154.2, 147.3, 146.7, 127.5, 127.3, 117.8, 79.6, 63.8, 61.5, 61.3, 61.0, 58.3, 58.2, 37.7, 28.1, 19.7, 14.3 ppm; HRMS (FAB) m/z [M+H]⁺ calcd for C₂₄H₂₉N₆O₆S₃: 593.1310; found: 593.1301.

Boc-alanylpiperidine 23: To a solution of 8 (262 mg, 4.42×10^{-1} mmol), DMAP (10.8 mg, 8.84×10^{-2} mmol), and NEt₃ (0.0678 mL, $4.86 \times$ 10^{-1} mmol) in dry THF (4.4 mL) at 0° C under Ar atmosphere was added Boc₂O (0.112 mL, 4.86×10^{-1} mmol). The solution was stirred at 0°C for 1 h. The reaction was quenched with H_2O (4 mL) and the mixture was extracted with AcOEt (4 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (80–90% AcOEt/hexane) to afford the Boc-oxazolidinone (257 mg, 84%) as colorless solids: $[a]_D^{27} + 60.2$ (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 1820, 1725, 1480, 1375, 1325, 1155, 1100, 1080, 1045,$ 1020 cm⁻¹; ¹H NMR (CDCl₃) δ = 8.15 (s, 1H, thiazole H-5), 7.96 (s, 1H, thiazole H-5), 6.97 (s, 1H, thiazole H-5), 5.01 (d, $J=4.5$ Hz, 1H, oxazolidinone H-4), 4.90 (s, 1H, piperidine H-6), 4.56 (dq, 1H, J=4.5, 6.6 Hz, oxazolidinone H-5), 4.52-4.36 (m, 5H, piperidine H-2, $CO_2CH_2CH_3\times2$), 2.78–2.62 (m, 1H, piperidine H-4b), 2.46–2.30 (m, 1H, piperidine), 2.16– 1.96 (m, 2H, piperidine), 1.52 (d, $J=6.6$ Hz, 3H, oxazolidinone 5-Me), 1.46–1.37 (m, 6H, CO₂CH₂CH₃×2), 1.42 ppm (s, 9H, Boc); ¹³C NMR (CDCl₃): δ = 181.6, 173.7, 166.9, 161.5, 161.4, 154.1, 150.8, 148.7, 147.4, 147.0, 127.6, 127.2, 117.7, 84.7, 75.8, 63.8, 62.4, 61.5, 61.2, 58.3, 58.1, 37.9, 27.9, 27.8, 20.0, 14.3 ppm; HRMS (FAB): m/z [$M+H$]⁺ calcd for $C_{29}H_{37}N_6O_8S_3$: 693.1835; found: 693.1839. To a solution of the above Bocoxazolidinone (245 mg, 3.54×10^{-1} mmol), N-Boc-L-alanine 22 (134 mg, 7.08×10^{-1} mmol), HOAt (134 mg, 7.08×10^{-1} mmol), and iPr_2NEt (0.278 mL, 1.59 mmol) in CH_2Cl_2 (3.5 mL) at room temperature under Ar atmosphere was added CIP (197 mg, 7.08×10^{-1} mmol). The solution was stirred for 1 day and quenched with $H₂O$ (4 mL). The mixture was extracted with AcOEt ($4 mL \times 3$). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was dissolved in CHCl₃ and this was filtered through celite and the filtrate was evaporated. The residue was chromatographed on silica gel (70–80% AcOEt/hexane) to afford 23 (284 mg, 93%) as colorless solids: $\lbrack a \rbrack_{D}^{30}$ +20.9 (c 1.00, MeOH); IR (CHCl₃): $\tilde{v} = 1820, 1795, 1720, 1500, 1370,$ 1325, 1160, 1100, 1070, 1045 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.54 (brs, 1H, amide NH), 8.12 (s, 1H, thiazole H-5), 7.84 (s, 1H, thiazole H-5), 6.89 (s, 1H, thiazole H-5), 5.45 (br d, $J=6.0$ Hz, 1H, BocNH), 5.17 (d, $J=$ 2.7 Hz, 1H, oxazolidinone H-4), 5.03 (dq, J=2.7, 6.3 Hz, 1H, oxazolidinone H-5), 4.59 (s, 1H, piperidine H-6), 4.46 (dd, J=2.7, 10.1 Hz, 1H, piperidine H-2), 4.41 (q, $J=7.2$ Hz, $2H$, $CO_2CH_2CH_3$), 4.40 (q, $J=7.2$ Hz, 2H, CO₂CH₂CH₃), 4.16 (dq, $J=6.0$, 6.6 Hz, 1H, Ala H- α), 3.51 (br d, $J=$ 14.1 Hz, 1 H, piperidine H-4 α), 2.72 (ddd, J = 14.1, 14.1, 3.4 Hz, 1 H, piperidine H-4 β), 2.24 (brd, J=12.0 Hz, 1H, piperidine H-3 β), 2.01 (br ddd, $J=12.0$, 14.1, 10.1 Hz, 1H, piperidine H-3 α), 1.61 (d, $J=6.3$ Hz, 3H, oxazolidinone 5-Me), 1.46 (s, 9H, Boc), 1.45 (d, J=6.6 Hz, 3H, Ala Me), 1.39 (t, $J=7.2$ Hz, 4H, CO₂CH₂CH₃×2), 1.30 ppm (s, 9H, Boc); ¹³C NMR (CDCl₃): δ =174.9, 173.3, 173.0, 167.4, 161.4, 161.3, 155.3, 152.8, 151.3, 148.9, 146.8, 127.3, 127.1, 119.7, 85.4, 79.7, 75.8, 64.9, 61.5, 61.4, 61.3, 61.2, 58.0, 51.4, 30.7, 28.2, 28.1, 27.8, 27.4, 20.4, 18.7, 14.3 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C₃₇H₅₀N₇O₁₁S₃: 864.2731; found: 864.2726.

Dehydropiperidine 25: To a solution of 23 (100 mg, 1.16×10^{-1} mmol) in THF (1.2 mL) at -78° C under Ar atmosphere was added *tert*-butyl hypochlorite (0.144 mL, 1.28×10^{-1} mmol). The solution was stirred for 30 min and then DMAP $(2.8 \text{ mg}, 2.3 \times 10^{-2} \text{ mmol})$ and NEt₃ $(0.162 \text{ mL},$ 1.16 mmol) were added. The temperature was raised to room temperature and the reaction mixture was stirred for 5 h. The reaction was quenched with saturated aqueous $Na₂S₂O₃$ (1 mL) and saturated aqueous NaHCO₃ (1 mL). The mixture was extracted with AcOEt (2 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (35–40% acetone/hexane) to afford 25 (94.6 mg, 95%) as pale yellow solids: $\lbrack \alpha \rbrack_{D}^{30}$

 $+23.3$ (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu} = 1820, 1720, 1500, 1370, 1330,$ 1160, 1100, 1070, 1045 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.43 (brs, 1H, amide NH), 8.19 (s, 1H, thiazole H-5), 7.91 (s, 1H, thiazole H-5), 7.00 (br s, 1H, thiazole H-5), 5.43 (br s, 1H, piperidine H-6), 5.30–5.12 (m, 3H, BocNH, oxazolidinone H-4, oxazolidinone H-5), 4.51-4.34 (m, 4H, $CO_2CH_2CH_3\times$ 2), 3.99 (dq, $J=7.2$, 5.7 Hz, 1H, Ala H- α), 3.63 (ddd, $J=13.8$, 5.4, 0.0 Hz, 1H, piperidine H-4a), 3.39 (m, 1H, piperidine H-3b), 3.12–2.92 (m, 1H, piperidine H-3 β), 2.80 (ddd, J = 13.8, 13.2, 6.0 Hz, 1H, piperidine H-4 β), 1.61(d, J=6.3 Hz, 3H, oxazolidinone 5-Me), 1.50 (s, 9H, Boc), 1.47–1.35 (m, 6H, CO₂CH₂CH₃×2), 1.35 (d, J=7.2 Hz, 3H, Ala Me), 1.21 ppm (s, 9H, Boc); ¹³C NMR (CDCl₃): δ = 175.2, 173.7, 168.9, 167.6, 163.6, 161.2, 161.1, 155.2, 153.2, 150.5, 148.6, 147.9, 147.0, 130.0, 127.3, 120.0, 85.2, 79.6, 74.9, 66.7, 61.6, 61.4, 61.3, 59.8, 51.9, 27.9, 27.8, 26.9, 24.5, 20.5, 17.9, 14.25, 14.22 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C₃₇H₄₈N₇O₁₁S₃: 862.2574; found: 862.2573.

Alanylpiperidine 24: To a solution of 23 (50 mg, 5.79×10^{-2} mmol) in MeOH (0.25 mL), 1,4-dioxane (0.25 mL), and H₂O (0.5 mL) at 0°C was added LiOH (6.9 mg, 2.9×10^{-1} mmol). The mixture was stirred at room temperature for 2 h. The mixture was quenched with 1m aqueous HCl (1 mL) at 0° C and extracted with AcOEt (1 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was dissolved in 1,4-dioxane (0.5 mL) and 1 M aqueous HCl (1 mL), and the mixture was stirred at room temperature for 1 h. The solvent was evaporated to afford the crude amino acid. This was dissolved in MeOH (0.1 mL). Acetone (5 mL) was added to the solution to form precipitates. The precipitates were filtered off to afford 24·3HCl $(30.2 \text{ mg}, 76\%)$ as solids: $[\alpha]_D^{30} + 45.6$ (c 1.21 (3HCl), 1 M aqueous HCl) [Ref. [10f]: $[\alpha]_D^{29}$ +45 (1 M aqueous HCl); Ref. [37]: $[\alpha]_D^{26}$ +44.7 (c 1.02, 1 M aqueous HCl)]; IR (KBr, HCl free): $\tilde{v} = 1685, 1580, 1380$ cm⁻¹ [Ref. [10 f]: IR (KBr): $\tilde{v} = 1684 \text{ cm}^{-1}$; Ref. [37]: IR (KBr): $\tilde{v} = 1690$, 1580, 1480, 1370 cm⁻¹]; ¹H NMR (D₂O, HCl salt): δ = 8.64 (s, 1H, thiazole H-5), 8.49 (s, 1H, thiazole H-5), 7.59 (s, 1H, thiazole H-5), 5.45 (s, 1H, piperidine H-6), 5.37 (br d, J=10.5 Hz, 1H, piperidine H-2), 4.94 (d, J=6.9 Hz, 1H, H₂NCH), 4.50 (q, $J=6.9$ Hz, 1H, Ala H- α), 4.38 (dq, $J=6.9$, 6.6 Hz, 1H, CHCH(OH)CH₃), 3.44 (br d, $J=14.7$ Hz, 1H, piperidine H-4 α), 2.94 (br dd, $J=14.7$, 12.6 Hz, 1H, piperidine H-4 β), 2.79 (br d, $J=12.6$ Hz, 1H, piperidine H-3 β), 2.47 (br ddd, $J=10.5$, 12.6, 12.6 Hz, 1H, piperidine H-3a), 1.77 (d, $J=6.9$ Hz, 3H, Ala Me), 1.32 ppm (d, $J=6.6$ Hz, 3H, CHCH(OH)C<u>H</u>₃) [Ref. [37]: ¹H NMR (D₂O, HCl salt): $\delta = 8.61$ (s, 1H), 8.46 (s, 1H), 7.59 (s, 1H), 5.48 (s, 1H), 5.40 (1H), 4.52 (q, J=8 Hz, 1H), 4.40 (dq, $J=6.5$, 6.5 Hz, 1H), 3.6–3.3 (1H), 3.2–2.2 (3H), 1.77 (d, $J=$ 8 Hz, 3H), 1.30 ppm (d, J=6.5 Hz, 3H)]; ¹³C NMR (D₂O, HCl salt): δ = 172.3, 171.5, 166.4, 166.2, 165.2, 164.9, 147.5, 147.2, 146.2, 132.5, 132.1, 126.5, 68.9, 62.6, 62.2, 58.4, 58.3, 51.3, 32.1, 26.3, 20.1, 18.1 ppm [Ref. [37]: ¹³C NMR (D₂O, HCl salt): δ = 172.3, 171.4, 166.4, 165.8, 165.2, 164.8, 147.4, 147.1, 146.0, 132.6, 132.1, 126.6, 68.9, 62.6, 62.2, 58.4, 58.4, 51.4, 32.2, 26.2, 20.2, 18.2 ppm].

Adduct 20: A solution of 14 (280 mg, 1.32 mmol), 15a (205 mg, 1.32 mmol), LiClO₄ (1.40 g, 13.2 mmol), and NEt₃ (1.84 mL, 13.2 mmol) in dry THF (11 mL) was stirred at room temperature for 5 h under Ar atmosphere to form 12a. The temperature was cooled to -40° C and then 11 (500 mg, 1.32 mmol) was added to the above solution. The mixture was stirred at -25°C for 1 day. The reaction mixture was quenched with H₂O (30 mL) and extracted with AcOEt (30 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (90–100% AcOEt/ hexane) to afford 20 (628 mg, 65%) as colorless solids and its diastereomer (115 mg, 6%) as colorless solids, and 11 (129 mg, 26%) was recovered. **20**: ¹H NMR (CDCl₃): $\delta = 8.30$ (s, 1H, thiazole H-5), 7.99 (s, 1H, thiazole H-5), 7.45 (d, $J=12.0$ Hz, 2H, toluene), 7.27 (d, $J=12.0$ Hz, 2H, toluene), 6.96 (s, $1H$, thiazole H-5), 6.30 (brs, $1H$, oxazolidinone NH), 5.72 (d, $J=12.0$ Hz, 1H, NHSO), 5.05 (d, $J=12.0$ Hz, 1H, CHNHSO), 4.75 (dd, $J=9.6$, 1.8 Hz, 1H, oxazolidinone H-4), 4.51 (dq, $J=9.6$, 9.6 Hz, 1H, oxazolidinone H-5), 4.48-4.34 (m, 4H, $CO_2CH_2CH_3\times2$), 3.20-3.06 (m, 2H, pyrrolidine H-3×2), 2.75–2.51 (m, 2H, pyrrolidine H-4×2), 2.42 (s, 3H, toluene), 1.51 (d, $J=9.6$ Hz, 3H, oxazolidinone 5-Me), 1.41 (t, $J=$ 10.5 Hz, 3H, CO₂CH₂CH₃), 1.39 ppm (t, $J=10.5$ Hz, 3H, CO₂CH₂CH₃); ¹³C NMR (CDCl₃): δ = 174.3, 172.1, 168.3, 163.0, 161.3, 160.8, 158.2, 154.4, 148.6, 146.7, 141.6, 141.1, 131.0, 129.7, 128.0, 125.4, 117.5, 86.0,

79.7, 61.7, 61.4, 60.9, 59.7, 36.8, 31.5, 21.4, 19.7, 14.3, 14.2 ppm. Diastereomer of 20: ¹H NMR (CDCl₃): $\delta = 8.38$ (s, 1H, thiazole H-5), 8.11 (s, 1H, thiazole H-5), 7.13 (d, $J=8.4$ Hz, 2H, toluene), 7.08 (d, $J=8.4$ Hz, 2H, toluene), 7.08 (s, 1H, thiazole H-5), 6.25 (br s, 1H, oxazolidinone NH), 5.96 (d, J=10.5 Hz, 1H, NHSO), 4.81 (d, J=10.5 Hz, 1H, CHNHSO), 4.52 (dd, J=6.0, 0.9 Hz, 1H, oxazolidinone H-4), 4.50–4.37 (m, 4H, $CO_2CH_2CH_3 \times 2$, 4.34 (dq, $J=6.0$, 6.0 Hz, 1H, oxazolidinone H-5), 3.07– 2.92 (m, 1H, pyrrolidine H-3), 2.62–2.25 (m, 3H, pyrrolidine H-3, H-4 \times 2), 2.38 (s, 3H, toluene), 1.47–1.36 ppm (m, 9H, oxazolidinone 5-Me, $CO₂CH₂CH₃×2$.

Aminopiperidine 26: To a solution of 20 (235 mg, 3.22×10^{-1} mmol) in dry EtOH (3.2 mL) at 0° C was added TFA (0.248 mL, 3.22 mmol). The solution was stirred at room temperature for 1 h. The reaction was quenched with aqueous NaHCO₃ (5 mL) at 0° C. The mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over Na2SO4, filtered through celite, and evaporated to afford the crude amine. This crude amine and AcOH (0.185 mL, 3.22 mmol) were dissolved in dry EtOH (1 mL) . To the mixture was added NaBH₃CN (60.8 mg, 9.67×10^{-1} mmol) in dry EtOH (2.2 mL) at 0°C. The reaction mixture was stirred at room temperature for 2 h. The reaction was quenched with aqueous NaHCO₃ (5 mL) at 0° C. The mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (90–100% AcOEt/hexane) to afford 26 (90.0 mg, 47%) as colorless solids: ¹H NMR (CDCl₃): δ = 8.15 (s, 1H, thiazole H-5), 7.97 (s, 1H, thiazole H-5), 6.98 (s, 1H, thiazole H-5), 6.31 (br s, 1H, oxazolidinone NH), 4.89 (s, 1H, piperidine H-6), 4.72 (dd, $J=6.0$, 2.0 Hz, 1H, oxazolidinone H-4), 4.53–4.34 (m, 6H, piperidine H-2, oxazolidinone H-5, $CO_2CH_2CH_3\times 2$), 2.78–2.64 (m, 1H, piperidine H-4a), 2.50–2.25 (m, 1H, piperidine H-3), 2.15–1.96 (m, 2H, piperidine H-4b, H-3), 1.49 (d, $J=6.0$ Hz, 3H, oxazolidinone 5-Me), 1.40 ppm (t, $J=7.0$ Hz, 6H, CO₂CH₃CH₃×2); ¹³C NMR (CDCl₃): δ = 181.2, 173.6, 169.1, 161.6, 161.4, 158.2, 154.5, 147.4, 147.0, 127.6, 127.2, 117.5, 79.8, 64.0, 61.6, 61.4, 60.9, 58.3, 58.1, 38.0, 28.1, 19.8, 14.3 ppm.

Boc-alanylpiperidine 27: To a solution of 26 (85.8 mg, 1.45×10^{-1} mmol), DMAP (1.8 mg, 1.4×10^{-2} mmol), and NEt₃ (0.0222 mL, 1.59×10^{-1} mmol) in dry THF (1.5 mL) at 0° C under Ar atmosphere was added Boc₂O (0.0366 mL, 1.59×10^{-1} mmol). The solution was stirred at 0°C for 1 h. The reaction was quenched with H_2O (2 mL) and the mixture was extracted with AcOEt ($2 mL \times 3$). The combined extracts were dried over Na2SO4, filtered through celite, and evaporated. The residue was chromatographed on silica gel (80–100% AcOEt/hexane) to afford the Boc-oxazolidinone (83.0 mg, 83%) as colorless solids: m.p. 156–158 °C; $\left[\alpha\right]_D^{27}$ -109.2 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 1820, 1720, 1080, 1045$ cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.15$ (s, 1H, thiazole H-5), 7.97 (s, 1H, thiazole H-5), 7.01 (s, 1H, thiazole H-5), 5.02 (d, J=3.9 Hz, 1H, oxazolidinone H-4), 4.93 (s, 1H, piperidine H-6), 4.54–4.34 (m, 6H, piperidine H-2, oxazolidinone H-5, $CO_2CH_2CH_3\times 2$), 2.82–2.64 (m, 1H, piperidine H-4a), 2.45– 2.27 (m, 1H, piperidine), 2.15–1.95 (m, 2H, piperidine), 1.49 (d, $J=$ 6.4 Hz, 3H, oxazolidinone 5-Me), 1.42 (s, 9H, Boc), 1.41 ppm (t, $J=$ 7.1 Hz, 6H, CO₂CH₂CH₃×2); ¹³C NMR (CDCl₃): δ = 181.4, 173.7, 167.2, 161.6, 161.4, 154.2, 150.8, 148.6, 147.6, 147.0, 127.5, 127.2, 117.4, 84.7, 75.7, 64.0, 62.4, 61.5, 61.3, 58.4, 58.0, 37.9, 28.0, 27.9, 20.2, 14.4 ppm. To a solution of the above Boc-oxazolidinone (76.1 mg, 1.10×10^{-1} mmol), N-Boc-L-alanine 22 (41.6 mg, 2.20×10^{-1} mmol), HOAt (41.6 mg, $2.20 \times$ 10^{-1} mmol), and iPr_2NEt (0.0861 mL, 4.94×10^{-1} mmol) in CH₂Cl₂ (1.1 mL) at room temperature under Ar atmosphere was added CIP $(61.2 \text{ mg}, 2.20 \times 10^{-1} \text{ mmol})$. The solution was stirred at room temperature for 1 day and quenched with $H₂O$ (2 mL). The mixture was extracted with AcOEt ($2 \text{ mL} \times 3$). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was dissolved in CHCl₃ and this was filtered through celite and the filtrate was evaporated. The residue was chromatographed on silica gel (80% AcOEt/hexane) to afford 27 (94.9 mg, quantitative yield): ¹H NMR (CDCl₃): $\delta = 8.15$ (s, 1H, thiazole H-5), 8.05 (br s, 1H, amide NH), 7.88 (s, 1H, thiazole H-5), 6.93 (s, 1H, thiazole H-5), 5.27 (brs, 1H, BocNH), 5.13 (d, $J=3.4$ Hz, 1H, oxazolidinone H-4), 4.59 (s, 1H, piperidine H-6), 4.49 (dd, $J=2.8$, 11.4 Hz, 1H, piperidine H-2), 4.49–4.22 (m, 6H, oxazolidinone H-5, Ala H- α , CO₂CH₂CH₃×2), 3.43 (brd, J=14.0 Hz, 1H, piperidine H-4 β),

2.86–2.71 (m, 1H, piperidine H-4 α), 2.34–2.19 (m, 1H, piperidine H-3 α), 2.07–1.85 (m, 1H, piperidine H-3 β), 1.58 (d, $J=6.0$ Hz, 3H, oxazolidinone 5-Me), 1.47 (s, 9H, Boc), 1.50–1.34 (m, 9H, Ala Me, $CO_2CH_2CH_3\times$ 2), 1.38 ppm (s, 9H, Boc).

Alanylpiperidine 28: To a solution of 27 (90.0 mg, 1.04×10^{-1} mmol) in MeOH (0.5 mL), 1,4-dioxane (0.5 mL), and H₂O (1 mL) at 0°C was added LiOH (6.9 mg, 5.2×10^{-1} mmol). The mixture was stirred at room temperature for 2 h. The mixture was quenched with 1m aqueous HCl (2 mL) at 0° C and extracted with AcOEt (2 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was dissolved in 1,4-dioxane (1 mL) and 1m aqueous HCl (2 mL), and the solution was stirred at room temperature for 1 h. The solvent was evaporated to afford the crude amino acid. This was dissolved in MeOH (0.1 mL). Acetone (5 mL) was added to the solution to form precipitates. The precipitates were filtered off to afford 28·3HCl $(50.6 \text{ mg}, 70\%)$ as solids: α_{D}^{30} -45.9 (c 1.21 (3HCl), 1 M aqueous HCl);
¹H NMP (D.O. HCl solt): $\lambda = 8.63$ (s 1 H this sole H 5), 8.40 (s 1 H this ¹H NMR (D₂O, HCl salt): δ = 8.63 (s, 1H, thiazole H-5), 8.40 (s, 1H, thiazole H-5), 7.50 (s, 1H, thiazole H-5), 5.39 (s, 1H, piperidine H-6), 5.36 (dd, $J=12.6$, 3.3 Hz, 1H, piperidine H-2), 4.88 (d, $J=7.5$ Hz, 1H, H₂NCH), 4.57 (q, $J=7.2$ Hz, 1H, Ala H- α), 4.46 (dq, $J=7.5$, 6.3 Hz, 1H, $CHCH(OH)CH₃$), 3.52 (brd, $J=14.1$ Hz, 1H, piperidine H-46), 3.05 (ddd, $J=14.1$, 14.1, 3.3 Hz, 1H, piperidine H-4 α), 2.75 (br d, $J=15.0$ Hz, 1H, piperidine H-3 α), 2.54 (br ddd, $J=15.0$, 14.1, 12.6 Hz, 1H, piperidine H-3 β), 1.75 (d, J=7.2 Hz, 3H, Ala Me), 1.35 ppm (d, J=6.3 Hz, 3H, CHCH(OH)CH₃); ¹³C NMR (D₂O, HCl salt): δ = 172.4, 171.9, 166.6, 166.5, 165.4, 165.1, 147.8, 147.6, 146.9, 132.3, 131.7, 126.4, 69.2, 63.3, 62.8, 58.5, 58.2, 50.7, 31.4, 26.2, 20.5, 17.7 ppm.

TMSE ester 29: To a solution of 8 (4.94 g, 8.33 mmol) in 2-(trimethylsilyl)ethanol (11.9 mL, 83.5 mol) was added $Ti(OiPr)₄$ (2.44 mL, 8.33 mmol). After stirring at 100 $^{\circ}$ C for 6 h, the reaction mixture was quenched with $H₂O$ (100 mL) and extracted with CHCl₃ (100 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (80% AcOEt/hexane) to afford 29 (4.61 g, 75%) as a colorless foam: $R_f = 0.63$ (100% AcOEt); $[\alpha]_D^{30}$ +95.4 (c 1.00, CHCl₃); m.p. 84–85 °C; IR (CHCl₃): $\tilde{\nu} = 3440, 3320$, 1760, 1720, 1480, 1390, 1340, 1100, 1040, 970 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.08 (s, 1H, thiazole H-5), 7.89 (s, 1H, thiazole H-5), 6.92 (s, 1H, thiazole H-5), 6.70 (br s, 1H, oxazolidinone NH), 4.82 (s, 1H, piperidine H-6), 4.71 (br d, J=6.3 Hz, 1H, oxazolidinone H-4), 4.51–4.34 (m, 6H, piperidine H-2, oxazolidinone H-5, and $Me₃SiCH₂CH₂ \times 2)$, 2.78–2.56 (m, 1H, piperidine H-4), 2.52–2.20 (m, 1H, piperidine H-3), 2.15–1.96 (m, 2H, piperidine H-3 and H-4), 1.50 (d, $J=6.3$ Hz, 3H, oxazolidinone 5-Me), 1.18-1.04 (m, 4H, Me₃SiCH₂CH₂×2), 0.06 (s, 9H, Me₃SiCH₂CH₂), 0.05 ppm (s, 9H, \underline{Me}_3 SiCH₂CH₂); ¹³C NMR(CDCl₃): $\delta = 181.47, 173.36,$ 169.22, 161.66, 161.40, 158.25, 154.37, 147.61, 147.13, 127.31, 126.94, 117.74, 79.72, 63.80, 63.73, 63.51, 61.01, 58.31, 58.09, 37.74, 27.81, 19.65, 17.43, 17.33, -1.47 , -1.53 ppm; HRMS (FAB): m/z [M-H]⁻ calcd for $C_{30}H_{43}N_6O_6S_3Si_2$: 735.1945; found: 735.1970.

Bpoc-alanylpiperidine 31: To a solution of 29 (5.31 g, 7.20 mmol), DMAP (176 mg, 1.44 mmol), and NEt₃ (1.50 mL, 10.8 mmol) in dry THF (72 mL) at 0° C under Ar atmosphere was added Boc₂O (1.66 mL, 7.23 mmol). After stirring at 0° C for 1.5 h, the reaction mixture was quenched with $H₂O$ (80 mL) and extracted with AcOEt (100 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (40% AcOEt/hexane) to afford the Boc-oxazolidinone (5.61 g, 93%) as a colorless foam: R_f = 0.80 (70% AcOEt/hexane); ¹H NMR (CDCl₃): $\delta = 8.10$ (s, 1H, thiazole H-5), 7.91 (s, 1H, thiazole H-5), 6.93 (s, 1H, thiazole H-5), 4.99 (d, J=4.7 Hz, 1H, oxazolidinone H-4), 4.88 (br d, J=6.3 Hz, 1H, piperidine H-2), 4.55 (dq, J=4.7, 6.6 Hz, 1H, oxazolidinone H-5), 4.49–4.36 (m, 5H, piperidine H-6, Me₃SiCH₂CH₂×2), 2.74–2.58 (m, 1H, piperidine H-4), 2.43–2.24 (m, 1H, piperidine H-3), 2.12–1.94 (m, 2H, piperidine H-3 and H-4), 1.50 (d, $J=6.6$ Hz, 3H, oxazolidinone 5-Me), 1.40 (s, 9H, Boc), 1.19–1.07 (m, 4H, $Me₃SiCH₂CH₂×2), 0.07$ (s, 9H, $Me₃SiCH₂CH₂), 0.06 ppm$ (s, 9H, $Me₃SiCH₂CH₂$). To a solution of this Boc-oxazolidinone (4.09 g, 4.89 mmol), Bpoc-l-Ala-OH (30) (3.20 g, 9.77 mmol), HOAt (1.33 g, 9.77 mmol), and iPr_2NEt (4.25 mL, 24.4 mmol) in dry CH₂Cl₂ (9.8 mL) at room temperature under Ar atmosphere was added CIP (2.72 g,

9.76 mmol). After stirring at room temperature for 3.5 h, the reaction mixture was quenched with $H₂O$ (20 mL) and the mixture was extracted with CHCl₃ (30 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (40% AcOEt/hexane) to afford 31 (4.70 g, 84%) as a colorless foam: $R_f = 0.56$ (60% AcOEt/hexane); $[a]_D^{32} + 29.6$ (c 1.00, CHCl₃); m.p. 100–102°C; IR (CHCl₃): $\tilde{v} = 3420, 3370, 1820, 1720, 1490, 1420,$ 1370, 1320, 1100, 1040 cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.47$ (brs, 1H, CONH), 8.05 (s, 1H, thiazole H-5), 7.68 (s, 1H, thiazole H-5), 7.56–7.23 (m, 9H, Bpoc), 6.74 (s, 1H, thiazole H-5), 5.52 (brd, $J=6.0$ Hz, 1H, NHBpoc), 5.06 (d, J=2.7 Hz, 1H, oxazolidinone H-4), 4.96–4.80 (m, 1H, oxazolidinone H-5), 4.48 (s, 1H, piperidine H-6), 4.46–4.31 (m, 5H, piperidine H-2, Me₃SiCH₂CH₂×2), 4.10 (dq, J=6.0, 6.3 Hz, 1H, Ala H-a), 3.46 (br d, J=13.8 Hz, 1H, piperidine H-4), 2.71 (ddd, J=3.3, 13.8, 13.8 Hz, 1H, piperidine H-4), 2.19 (br dd, J=2.7, 13.8 Hz, 1H, piperidine H-3), 2.08–1.87 (m, 1H, piperidine H-3), 1.59 (d, J=4.2 Hz, 3H, oxazolidinone 5-Me), 1.41 (s, 9H, Boc), 1.35 (d, $J=6.3$ Hz, 3H, Ala Me), 1.16– 1.04 (m, 4H, Me₃SiC<u>H</u>₂CH₂×2), 0.05 (s, 9H, <u>Me₃SiCH₂CH₂), 0.03 ppm (s</u>, 9H, Me_3 SiCH₂CH₂); ¹³C NMR(CDCl₃): δ = 174.69, 172.96, 172.81, 168.01, 161.25, 161.19, 154.53, 152.79, 150.60, 148.49, 146.96, 146.85, 145.12, 140.46, 139.43, 128.59, 127.10, 127.04, 126.82, 126.70, 124.66, 119.19, 84.85, 80.77, 75.50, 65.23, 63.60, 63.39, 61.48, 61.17, 58.06, 51.48, 30.51, 29.42, 27.73, 27.15, 20.30, 18.16, 17.34, 17.22, -1.55, -1.63 ppm; HRMS (FAB): m/z $[M+Na]^+$ calcd for $C_{54}H_{71}N_7O_{11}S_3S_1Na$: 1168.3810; found: 1168.3782. Bpoc-l-Ala-OH 30 was prepapred as follows: To a solution of 4-acetylbiphenyl (15.0 g, 7.64×10^{-2} mol) in dry Et₂O (380 mL) at 0[°]C under Ar atmosphere was added 3m MeMgBr in Et₂O (56.1 mL, $1.68 \times$ 10^{-1} mol). After stirring at room temperature for 1 h, the reaction mixture was quenched with saturated aqueous NH₄Cl (500 mL) and H_2O (250 mL), and the mixture was extracted with $Et₂O$ (500 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (30% AcOEt/hexane) to afford 1-(4-biphenylyl)-1-methylethanol (14.6 g, 90%) as colorless solids: ¹H NMR (CDCl₃): δ = 7.63–7.30 (m, 9H, biphenyl), 1.62 ppm (s, $6H$, Me \times 2). To a solution of 1-(4-biphenylyl)-1-methylethanol (4.14 g, 19.5 mmol) and dry pyridine (2.33 mL, 28.8 mmol) in dry CH_2Cl_2 (20 mL) at -5° C under Ar atmosphere was added dropwise ClCO₂Ph (2.94 mL, 23.4 mmol) in dry CH_2Cl_2 (10 mL) over 0.5 h. The reaction mixture was stirred at 0° C for 1 day and quenched with ice-cold water (10 mL) and CH_2Cl_2 (20 mL) was added. The mixture was washed with H₂O (40 mL \times 3). The organic layers were dried over Na₂SO₄, filtered through celite, and evaporated. The residual solid was recrystallized from AcOEt to afford 1-(4-biphenylyl)-1-methylethyl phenyl carbonate $(5.65 \text{ g}, 87\%)$ as colorless solids: ¹H NMR (CDCl₃) $\delta = 7.63 - 7.23$ (m, 14H, biphenyl and Ph), 1.63 ppm (s, 6H, $Me \times 2$). L-alanine (1.02 g, 11.4 mmol) was dissolved in 40 wt% Triton B in MeOH (5.6 mL, 12.3 mmol) and the solvent was evaporated under reduced pressure. The residue was then evaporated twice more with DMF (17 mL) at 45° C under vacuum in order to remove traces of water. The remaining syrup was treated with 1-(4-biphenylyl)-1-methylethyl phenyl carbonate (4.10 g, 12.3 mmol) in DMF (17 mL) at 50 $^{\circ}$ C for 2 h. The reaction mixture was quenched with H₂O (20 mL) at 0^oC and washed with Et₂O (20 mL \times 3). The aqueous layer was acidified with cold 1_M aqueous citric acid to about pH 3 at 0° C and extracted with Et₂O (20 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated to afford Bpoc-L-Ala-OH 30 (2.50 g, 67%) as a colorless syrup: $R_f = 0.74$ $(15/55/65 \text{ H}_2\text{O/MeOH/CHCl}_3);$ ¹H NMR $(CDCl_3):$ $\delta = 10.80-9.80$ (brs, 1H, CO₂H), 7.62–7.26 (m, 9H, biphenyl), 5.31 (brd, $J=7.4$ Hz, 1H, BpocNH), 4.28 (br dq, J=6.8, 7.4 Hz, 1H, Ala H-a), 1.79 (br s, 6H, Bpoc $Me \times 2$), 1.38 ppm (d, $J=6.8$ Hz, 1H, Ala Me).

Segment A (5) : To a solution of 31 (805 mg, 7.02×10^{-1} mmol) in 2-(trimethylsilyl)ethanol (7.0 mL) at 0° C under Ar atmosphere was added Cs_2CO_3 (229 mg, 7.03×10^{-1} mmol). After stirring at room temperature for 10 h, the reaction mixture was quenched with saturated aqueous NH₄Cl (5 mL) and H₂O (10 mL), and the mixture was extracted with AcOEt (20 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was distilled under reduced pressure (15 mmHg, 59°C) to recover 2-(trimethylsilyl)ethanol. The residue was chromatographed on silica gel (40% AcOEt/hexane) to afford the NHBoc-alcohol (629 mg, 80%) as a colorless foam: $R_f = 0.73$ (60% AcOEt/hexane); ¹H NMR (CDCl₃): $\delta = 8.56$ (brs, 1H, piperidine 5-CONH), 8.07 (s, 1H, thiazole H-5), 7.62–7.23 (m, 10H, biphenyl and thiazole H-5), 6.49 (s, 1H, thiazole H-5), 5.58 (brd, $J=9.3$ Hz, 1H, NHBoc), 5.43 (br d, J = 8.1 Hz, 1H, NHBpoc), 4.81 (br d, J = 9.3 Hz, 1H, Thr H-a), 4.77–4.64 (m, 1H, Thr H-b), 4.52–4.25 (m, 6H, piperidine H-2, H-6, and Me₃SiCH₂CH₂×2), 4.14 (brdq, $J=8.1$, 8.4 Hz, 1H, Ala H- α), 3.61 (br d, $J=3.9$ Hz, 1H, Thr β -OH), 3.38 (br d, $J=14.4$ Hz, 1H, piperidine H-4), 2.82–2.66 (m, 1H, piperidine H-4), 2.59–2.44 (br s, 1H, piperidine H-3), 2.24 (br d, $J=12.0$ Hz, 1H, piperidine H-3), 1.75 (s, 3H, Bpoc Me), 1.68 (s, 3H, Bpoc Me), 1.54–1.34 (m, 3H, Thr Me), 1.44 (s, 9H, Boc), 1.18–0.98 (m, 4H, $Me₃SiCH₂CH₂×2$), 1.14 (d, $J=8.4$ Hz, 3H, Ala Me), 0.07 (s, 9H, \underline{Me}_3 SiCH₂CH₂), 0.05 ppm (s, 9H, \underline{Me}_3 SiCH₂CH₂). To a solution of this NHBoc-alcohol (429 mg, 3.83×10^{-1} mmol) in dry THF (3.8 mL) at -78°C under Ar atmosphere was added *tert*-butyl hypochlorite (0.0476 mL, 0.421 mmol). The reaction mixture was stirred at -78° C for 1 h and then DMAP (9.4 mg, 7.7×10^{-2} mmol) and NEt₃ (0.533 mL, 3.83 mmol) were added. The temperature was raised to room temperature and the reaction mixture was stirred for 3 h. The reaction mixture was quenched with saturated aqueous $\text{Na}_2\text{S}_2\text{O}_3$ (2 mL) and saturated aqueous $NaHCO₃$ (4 mL), and the mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (30% AcOEt/hexane) to afford 5 (324 mg, 76%) as a pale yellow foam: $R_{\rm f} = 0.45$ (60% AcOEt/hexane); $[\alpha]_{\rm D}^{32}$ +47.9 (c 1.00, CHCl₃); m.p. 102–103 °C; IR (CHCl₃): $\tilde{v} = 3430, 3300, 1710, 1500, 1370,$ 1100, 1040 cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.61$ (brs, 1H, piperidine 5-CONH), 8.17 (s, 1H, thiazole H-5), 7.62–7.28 (m, 10H, biphenyl and thiazole H-5), 6.63 (s, 1H, thiazole H-5), 5.66 (brd, $J=9.6$ Hz, 1H, NHBoc), 5.40 (br d, J=7.5 Hz, 1H, NHBpoc), 5.20 (br s, 1H, piperidine H-6), 4.86 (brd, $J=9.3$ Hz, 1H, Thr H- α), 4.63–4.52 (m, 1H, Thr H- β), 4.52–4.25 (m, 4H, Me₃SiCH₂CH₂×2), 3.97 (br dq, J=6.6, 7.5 Hz, 1H, Ala H- α), 3.74–3.43 (m, 2H, piperidine H-3 and H-4), 3.35 (brd, $J=13.2$ Hz, 1H, piperidine H-3), 2.81 (brd, $J=12.0$ Hz, 1H, piperidine H-4), 1.73 (s, 3H, Bpoc Me), 1.65 (s, 3H, Bpoc Me), 1.47 (s, 9H, Boc), 1.35 (d, $J=6.6$ Hz, 3H, Ala Me), 1.21-1.00 (m, 7H, Thr Me and $Me₃SiCH₂CH₂×2)$, 0.09 (s, 9H, Me₃SiCH₂CH₂), 0.08 ppm (s, 9H, Me₃SiCH₂CH₂); ¹³C NMR(CDCl₃): δ = 175.91, 175.02, 173.56, 169.13, 162.90, 161.39, 161.31, 155.82, 154.87, 152.75, 148.01, 147.03, 144.74, 140.49, 139.62, 130.26, 128.75, 127.48, 127.22, 126.93, 124.68, 117.48, 81.54, 80.20, 68.33, 66.66, 63.77, 63.43, 59.95, 57.75, 51.36, 28.95, 28.52, 28.29, 26.40, 24.60, 20.15, 18.70, 17.40, 17.33, -1.48 , -1.55 ppm; HRMS (FAB): m/z $[M + Na]$ ⁺ calcd for $C_{53}H_{71}N_7O_{10}S_3Si_2Na$: 1140.3861; found: 1140.3877.

 β -Lactone 42: To a solution of L-threonine 18 (2.00 g, 16.8 mmol) in H₂O (8.4 mL) and 1,4-dioxane (8.4 mL) at room temperature were added NEt₃ (7.00 mL, 50.4 mmol) and TeocOPh(p -NO₂) (5.23 g, 18.5 mmol). After stirring at room temperature for 21 h, the reaction mixture was quenched with saturated aqueous $NaHCO₃$ (10 mL) and the mixture was washed with Et₂O (50 mL \times 1). The aqueous layer was acidified with 1 M aqueous HCl to pH 3 at 0° C and extracted with AcOEt (100 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (70% AcOEt/CHCl₃) to afford the Teoc-L-Thr-OH $(4.51 g,$ quantitative yield) as a colorless syrup: $R_f = 0.50$ (70% AcOEt/CHCl₃); ¹H NMR (CDCl₃): δ = 5.94–5.76 (m, 1H, TeocNH), 4.56–4.27 (m, 2H, H- α and H- β), 4.27– 4.02 (m, 2H, Me₃SiCH₂CH₂), 1.24 (d, 3H, $J=6.2$ Hz, Me- β), 1.00 (t, 2H, $J=8.6$ Hz, Me₃SiCH₂CH₂), 0.02 ppm (s, 9H, Me₃SiCH₂CH₂). To a solution of the Teoc-L-Thr-OH (50.0 mg, 2.14×10^{-1} mmol) in acetone (1.9 mL) at 0° C under Ar atmosphere were added acetone dimethylacetal (0.0700 mL, 5.67×10^{-1} mmol) and p-TsOH (3.3 mg, 1.9×10^{-2} mmol). After stirring at room temperature for 6 h, the reaction mixture was quenched with saturated aqueous $NaHCO₃$ (10 mL) and then NaCl (solid) was added. The mixture was extracted with AcOEt (10 mL \times 3). The combined extracts were dried over Na2SO4, filtered through celite, and evaporated to afford acetonide 35 (65.0 mg, quantitative yield) as a colorless syrup: $R_f = 0.90$ (70% AcOEt/CHCl₃); ¹H NMR (CDCl₃): $\delta =$ 4.30–3.96 (m, 3H, Me₃SiCH₂CH₂ and H- β), 3.94–3.74 (m, 1H, H- α), 1.55 (s, 6H, oxazolidine 2-Me), 1.38 (d, $J = 5.0$ Hz, 3H, Me- β), 1.00 (brt, $J =$ 8.2 Hz, 2H, Me₃SiCH₂CH₂), 0.02 ppm (s, 9H, Me₃SiCH₂CH₂). To a solu-

tion of 35 (50.0 mg, 1.65×10^{-1} mmol) and 36^[47] (49.5 mg, $1.81 \times$ 10^{-1} mmol) in dry DMF (1.5 mL) at 0°C under Ar atmosphere were added iPr_2NEt (71.8 mL, 4.12×10^{-1} mmol) and PyBOP (103 mg, 1.98 \times 10^{-1} mmol). After stirring at room temperature for 3.5 h, the reaction mixture was quenched with saturated aqueous $NaHCO₃$ (5 mL) and the mixture was extracted with AcOEt (10 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (20% AcOEt/hexane) to afford 42 (42.1 mg, 66%) as a colorless foam: $R_f = 0.80$ (5% MeOH/CHCl₃); $[\alpha]_D^{26}$ –4.78 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu}$ = 3682, 3620, 1832, 1698, 1520, 1478, 1420, 1456, 1048 cm⁻¹; ¹H NMR (CDCl₃): δ = 7.56–6.64 (m, 1H, CONH), 5.60 (dd, $J=6.0$, 8.0 Hz, 1H, β -lactone H- α), 4.90 (dq, $J=6.0$, 6.0 Hz, 1H, b-lactone H-b), 4.38–4.10 (m, 1H, oxazolidine H-5), 4.16 (t, $J=8.8$ Hz, 2H, Me₃SiCH₂CH₂), 3.90 (d, $J=7.0$ Hz, 1H, oxazolidine H-4), 1.63 (s, 3H, oxazolidine 2-Me), 1.57 (s, 3H, oxazolidine 2-Me), 1.45 (d, $J=7.0$ Hz, 3H, oxazolidine 5-Me), 1.40 (d, $J=6.0$ Hz, 3H, β -lactone Me), 1.10–0.90 (m, 2H, Me₃SiCH₂CH₂), 0.02 ppm (s, 9H, Me₃SiCH₂CH₂); ¹³C NMR (CDCl₃): δ = 169.94, 168.81, 153.21(br), 95.29, 74.85, 74.02, 67.00, 64.22, 58.77, 27.31, 25.43, 19.01, 17.83, 15.02, -1.61 ppm. Elemental analysis (%) calcd for $C_{17}H_{30}N_2O_6Si$: C 52.83, H 7.82, N 7.25; found: C 52.76, H 7.82, N 7.23.

 β -Lactone 43: To a solution of 36 (9.15 g, 33.5 mmol) in dry THF (335 mL) were added iPr_2NEt (7.01 mL, 40.2 mmol) and Boc₂O (15.4 mL, 67.0 mmol). The mixture was stirred at room temperature for 17 h. The reaction mixture was quenched with $H₂O$ (300 mL) and the mixture was extracted with AcOEt (300 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (10% AcOEt/hexane) to afford **43** (5.22 g, 77%): $[\alpha]_D^{27}$ +29.4 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 1830$, 1720, 1390, 1370, 1340, 1040 cm⁻¹; ¹H NMR (CDCl₃): δ = 5.43 (dd, J = 6.2, 8.0 Hz, 1 H, H- α), 5.30 (brd, $J=8.0$ Hz, 1 H, BocNH), 4.86 (dq, $J=6.2$, 6.2 Hz, 1 H, H- β), 1.46 (s, 9 H, Boc), 1.49–1.43 ppm (d, $J = 6.2$ Hz, 3 H, Me); ¹³C NMR (CDCl₃): δ = 170.0, 155.0, 81.5, 75.4, 60.4, 28.5, 15.3 ppm. Elemental analysis (%) calcd for $C_9H_{15}NO_4$: C 53.72, H 7.51, N 6.96; found: C 53.74, H 7.58, N 6.95.

Phenylselenobutanoic acid 44: To a solution of 43 (4.35 g, 21.6 mmol) in degassed dry DMF (43 mL) at room temperature under Ar atmosphere was added benzeneselenol (3.44 mL, 32.4 mmol). The mixture was stirred at 80° C for 2 h. The reaction mixture was quenched with 1 M aqueous NaOH (43 mL) and H₂O (85 mL), and the mixture was washed with Et₂O (100 mL \times 3). The aqueous layer was acidified with 1 m aqueous HCl to pH 3 and extracted with AcOEt (100 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (0-5% MeOH/CHCl₃) to afford **44** (7.75 g, quantitative yield): $[\alpha]_D^{27}$ +23.8 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 1715$, 1500, 1370, 1160, 1080, 1040 cm⁻¹; ¹H NMR (CDCl₃): $\delta = 7.65-$ 7.54 (m, 2H, Ph), 7.36–7.23 (m, 3H, Ph), 6.10 (br s, 0.3H, BocNH), 5.24 $(\text{br d}, J=7.8 \text{ Hz}, 0.7 \text{ H}, \text{BocNH})$, 4.51 (brs, 1H, H- α), 3.68 (m, 1H, H- β), 1.52–1.37 (m, 3H, Me- β), 1.44 ppm (s, 9H, Boc); ¹³C NMR (CDCl₃): δ = 174.7, 155.5, 135.3, 129.2, 128.2, 128.1, 80.5, 57.9, 40.6, 28.2, 18.1 ppm; HRMS (EI): m/z [M]⁺ calcd for C₁₅H₂₁NO₄⁸⁰Se: 359.0636; found: 359.0629.

Tripeptide 46: To a solution of 42 (42.1 mg, 1.09×10^{-1} mmol) in degassed dry DMF (0.22 mL) at room temperature under Ar atmosphere was added benzeneselenol (0.0174 mL, 1.64×10^{-1} mmol). After stirring at 80°C for 2 h, the reaction mixture was quenched with 1 M aqueous NaOH (0.25 mL) and H₂O (0.50 mL), and the mixture was washed with Et_2O (1.0 mL \times 3). The aqueous layer was acidified with 1m aqueous HCl to pH 3 at 0° C and extracted with AcOEt (1.0 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (70% AcOEt/hexane) to afford 45 (55.7 mg, 94%) as a pale yellow foam: 1 H NMR (CDCl₃): δ = 7.60–7.48 (m, 2H, PhSe), 7.32–7.18 (m, 3H, PhSe), 6.88–6.38 (m, 1H, CONH), 4.82–4.68 (m, 1H, phenylselenoamino acid H-a), 4.28–3.98 (m, 3H, Me₃SiCH₂CH₂ and oxazolidine H-5), 3.83 (d, $J=7.0$ Hz, 1H, oxazolidine H-4), 3.74–3.60 (m, 1H, phenylselenoamino acid H-β), 1.62 (s, 3H, oxazolidine 2-Me), 1.57 (s, 3H, oxazolidine 2-Me), 1.45 (d, J=6.8 Hz, 3H, Me), 1.36 (d, $J=6.0$ Hz, 3H, Me), 1.04–0.88 (m, 2H, Me₃SiCH₂CH₂), 0.02 ppm (s, 9H, Me₃SiCH₂CH₂). To a solution of 45 (55.7 mg, $1.02 \times$ 10^{-1} mmol) and D-Ser-OMe·HCl 37 (17.5 mg, 1.12×10^{-1} mmol) in MeOH (1.0 mL) at 0 °C were added NMM $(0.0270 \text{ mL}, 2.46 \times 10^{-1} \text{ mmol})$ and DMTMM (34.0 mg, 1.23×10^{-1} mmol). After stirring at room temperature for 6 h, the reaction mixture was quenched with H_2O (2 mL) and the mixture was extracted with AcOEt $(2 mL \times 3)$. The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (50% AcOEt/hexane) to afford 46 (54.4 mg, 82%) as a colorless foam: $R_f = 0.80$ (10% MeOH/CHCl₃); $[\alpha]_{\text{D}}^{27}$ –77.7 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu}$ = 3685, 3620, 3480, 3370, 1748, 1660, 1508, 1468, 1382, 1355, 1080, 1046 cm⁻¹; ¹H NMR (CDCl₃): δ = 7.66–7.42 (m, 3H, CONH, PhSe), 7.36–7.21 (m, 3H, PhSe), 6.56–6.38 (br d, J=7.2 Hz, 1H, CONH), 4.58 (ddd, J=3.0, 5.0, 8.0 Hz, 1H, Ser Ha), 4.49 (dd, J=4.0, 7.2 Hz, 1H, phenylselenoamino acid H-a), 4.36–4.12 (m, 3H, Me₃SiCH₂CH₂ and oxazolidine H-5), 4.10-3.82 (m, 3H, phenylselenoamino acid H- β and Ser H- $\beta \times 2$), 3.92 (d, J=9.0 Hz, 1H, oxazolidine H-4), 1.70 (s, 3H, oxazolidine 2-Me), 1.66 (s, 3H, oxazolidine 2-Me), 1.46 (d, $J=6.0$ Hz, 3H, Me), 1.42 (d, $J=7.0$ Hz, 3H, Me), 1.04–0.88 (m, 2H, Me₃SiCH₂CH₂), 0.02 ppm (s, 9H, <u>Me</u>₃SiCH₂CH₂); ¹³C NMR (CDCl₃): δ = 170.43, 169.12, 168.59, 154.79, 134.74, 134.67, 129.36, 128.29, 127.73, 95.09, 74.22, 68.00, 65.88, 62.95, 56.13, 55.12, 52.28, 39.46, 27.37, 26.02, 18.41, 17.85, 16.30, -1.70 ppm. Elemental analysis (%) calcd for $C_{27}H_{43}N_3O_8^{80}$ SeSi: C 50.30, H 6.72, N 6.52; found: C 50.20, H 7.04, N 6.56. Carboxylic acid 33: To a solution of 46 (1.00 g, 1.55 mmol) in MeOH (5.0 mL) , H₂O (5.0 mL) , and 1,4-dioxane (5.0 mL) at 0° C was added 1 M aqueous NaOH (2.3 mL). After stirring at room temperature for 1 h, the reaction mixture was acidified with 1 μ aqueous HCl to pH 3 at 0 °C and extracted with AcOEt (20 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated to afford the crude 33 as a pale yellow foam, which was used for the next reaction without purification.

Olefin 48: A solution of 40 (1.53 g, 8.25 mmol), 41 (1.00 g, 8.25 mmol), and Cs_2CO_3 (2.69 g, 8.25 mmol) in dry CH_2Cl_2 (45 mL) was stirred at room temperature for 2 h. The mixture was filtered through celite and evaporated. The residue was chromatographed on silica gel (30% AcOEt/hexane) to afford tert-butylsulfinamide 39 (2.50 g, quantitative yield) as a yellow syrup: $R_f = 0.80$ (60% AcOEt/hexane); ¹H NMR (CDCl₃): $\delta = 8.86$ (d, $J = 1.2$ Hz, 1H, imine-H), 8.38 (d, $J = 1.2$ Hz, 1H, thiazole H-5), 4.48 (q, $J=7.0$ Hz, $2H$, $CO_2CH_2CH_3$), 1.42 (t, $J=7.0$ Hz, 3H, $CO_2CH_2CH_3$), 1.28 ppm (s, 9H, tBu). To a solution of (Z)-2-bromo-2-butene (1.32 mL, 13.0 mmol) in dry THF (19 mL) at -78° C under Ar atmosphere was added 1.62m tBuLi in pentane (16.0 mL, 26.0 mmol). After 5 min at -78 °C, 1.0m ZnCl₂ in ether (13.0mL, 13.0 mmol) was added and the temperature was raised to 0° C during a period of 15 min. The solution was cooled to -78° C and 39 (749 mg, 2.60 mmol) in dry THF (5.0 mL) was added and the temperature was raised to -40° C. After stirring at -40° C for 6 h, the reaction mixture was quenched with saturated aqueous NH4Cl (40 mL) and the mixture was extracted with Et₂O (50 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (55% AcOEt/CHCl₃) to afford **48** (778 mg, 87%) as a colorless syrup: $R_f = 0.40$ (60% AcOEt/CHCl₃); $[\alpha]_D^{25}$ +144 (c 1.00, CHCl₃); IR (neat): $\tilde{v} = 3450, 3255, 3195, 3120, 2978, 1724, 1478, 1370, 1340, 1320,$ 1238, 1210, 1062, 1020, 960, 900, 880, 756 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.13 (s, 1H, thiazole H-5), 5.79 (brd, 1H, $tBuS(O)NH$), 5.74 (q, J= 7.0 Hz, 1 H, H- γ), 4.80 (brd, 1 H, H- α), 4.39 (q, J = 7.0 Hz, 2 H, CO₂CH₂CH₃), 1.89 (d, J = 7.0 Hz, 3H, Me- γ), 1.61 (s, 3H, Me- β), 1.40 (t, $J=7.0$ Hz, 3H, CO₂CH₂CH₃), 1.31 ppm (s, 9H, Boc); ¹³C NMR (CDCl₃): δ = 171.17, 160.98, 146.68, 132.39, 127.93, 127.90, 61.26, 55.75, 54.51, 22.57, 18.13, 14.18, 13.42 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for $C_{15}H_{25}N_2O_3S_2$: 345.1304; found: 345.1307.

NHBoc olefin 38: To a solution of 48 (778 mg, 2.26 mmol) in MeOH (15 mL) at 0° C was added 10% HCl/MeOH (15 mL). After stirring at room temperature for 0.5 h, the reaction mixture was evaporated and to the residue was added saturated aqueous $NaHCO₃$ (20 mL). This was extracted with AcOEt (20 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated to afford the amine as a pale yellow oil. To a solution of this amine in 1,4-dioxane (22 mL) under

Ar atmosphere were added NEt₃ (0.378 mL, 2.71 mmol) and Boc₂O (0.651 mL, 2.71 mmol). After stiring at room temperature for 2 h, the reaction mixture was quenched with saturated aqueous NaHCO_3 (30 mL) and the mixture was extracted with AcOEt (30 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (10% AcOEt/hexane) to afford 38 (635 mg, 83% from 48) as a colorless oil: $R_f = 0.90$ (70%) AcOEt/CHCl₃); $[\alpha]_D^{26}$ +80.0 (c 1.00, CHCl₃); IR (neat): $\tilde{\nu} = 3425$, 3355, 2975, 2938, 1720, 1500, 1368, 1322, 1238, 1210, 1166, 1098, 1060, 1016, 958, 880, 756 cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.11$ (s, 1H, thiazole H-5), 6.06–5.74 (m, 2H, BocNH and H- α), 5.56 (q, J=7.0 Hz, 1H, H- γ), 4.41 (q, J = 7.0 Hz, 2H, CO₂CH₂CH₃), 1.88 (d, J = 7.0 Hz, 3H, Me- γ), 1.59 (s, 3H, Me-β), 1.46 (s, 9H, Boc), 1.41 ppm (t, J=7.0 Hz, 3H, CO₂CH₂CH₃); ¹³C NMR (CDCl₃): δ = 171.37, 161.16, 154.91, 146.68, 134.25, 127.86, 124.62, 79.93, 61.41, 52.59, 28.34, 18.11, 14.30, 13.47 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C₁₆H₂₅N₂O₄S: 340.1457; found: 340.1457.

Diol 49: To a solution of OsO₄ (47.4 mg, 1.86×10^{-1} mmol) in tBuOH/ H₂O (85:15, 6.0 mL) were added 38 (635 mg, 1.87 mmol) in t BuOH/H₂O (85:15, 8.0 mL), NMO (656 mg, 5.60 mmol) in tBuOH/H2O (85:15, 4.0 mL), and DABCO (41.9 mg, 3.74×10^{-1} mmol). The reaction mixture was stirred in the dark at room temperature for 12 h and $Na₂S₂O₃$ was added. The resulting solution was stirred at room temperature for 1.5 h and then H₂O (20 mL) was added. The mixture was extracted with AcOEt (30 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (35% AcOEt/hexane) to afford 49 (390 mg, 56%) as a colorless foam and its diastereomer (175 mg, 25%) as a colorless foam. 49: R_f =0.45 (50% AcOEt/hexane); $[\alpha]_D^{29}$ -46.5 (c 1.00, CHCl₃); IR (neat): $\tilde{v} = 3420, 2980, 1720, 1498, 1370, 1340, 1326, 1220, 1164, 1100, 1016, 876,$ 756 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.12 (s, 1H, thiazole H-5), 6.02 (d, J = 9.0 Hz, 1 H, BocNH), 5.20 (d, $J=9.0$ Hz, 1 H, H- α), 4.38 (q, $J=7.0$ Hz, 2H, CO₂CH₂CH₃), 4.30–4.16 (m, 1H, OH), 3.84–3.70 (m, 1H, H-γ), 3.16– 3.02 (m, 1H, OH), 1.45 (s, 9H, Boc), 1.39 (t, J=7.0 Hz, 3H, CO₂CH₂CH₃), 1.24 (d, J=6.4 Hz, 3H, Me- γ), 1.12 ppm (s, 3H, Me- β); ¹³C NMR (CDCl₃): δ = 170.02, 160.81, 156.58, 146.99, 127.04, 80.54, 69.35, 61.34, 57.19, 28.28, 18.46, 16.70, 14.21 ppm; HRMS (FAB): m/z [$M+H$]⁺ calcd for $C_{16}H_{27}N_2O_6S$: 375.1591; found: 375.1590. Diastereomer of 49: $R_{\rm f}$ =0.30 (50% AcOEt/hexane); $\left[\alpha_{\rm D}^{28}$ -39.0 (c 1.00, CHCl₃); IR (neat): n˜ =3420, 2980, 1720, 1498, 1370, 1340, 1326, 1220, 1164, 1100, 1016, 876, 756 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.12 (s, 1H, thiazole H-5), 5.81 (d, J = 9.2 Hz, 1 H, BocNH), 5.20 (d, $J=9.2$ Hz, 1 H, H- α), 4.38 (q, $J=7.0$ Hz, 2H, CO₂CH₂CH₃), 3.68 (d, $J=8.0$ Hz, 1H, OH), 3.84–3.70 (dq, $J=7.0$, 8.0 Hz, 1 H, H- γ), 3.49 (s, 1 H, OH), 1.45 (s, 9 H, Boc), 1.39 (t, $J = 7.0$ Hz, 3H, CO₂CH₂CH₃), 1.32 (d, $J=6.4$ Hz, 3H, Me-y), 1.25 ppm (s, 3H, Me- β); ¹³C NMR (CDCl₃): $\delta = 171.15, 160.76, 155.40, 146.11, 127.27, 80.04,$ 71.23, 69.35, 61.13, 56.42, 28.00, 18.64, 16.89, 13.95 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C₁₆H₂₇N₂O₆S: 375.1591; found: 375.1590.

Aminodiol 50: To a solution of 49 (50.0 mg, 1.34×10^{-1} mmol) in EtOH (1.0 mL) and 1,4-dioxane (0.5 mL) at 0° C was added 1 M aqueous NaOH (0.2 mL). After stirring at room temperature for 0.5 h, the reaction mixture was acidified with 1_M aqueous HCl to pH 3 at 0° C and extracted with AcOEt (3 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was purified using preparative TLC (ODS, 50% MeOH/H₂O) to afford carboxylic acid (35.5 mg, 77%) as a colorless syrup: $R_f = 0.45$ (50% MeOH/H₂O, ODS); ¹H NMR (CD₃OD): $\delta = 8.24$ (brs, 1H, thiazole H-5), 6.96 (brd, J= 8.0 Hz, 1H, BocNH), 5.18 (br s, 1H, H-a), 3.77 (br q, J=5.8 Hz, 1H, H- γ), 1.43 (s, 9H, Boc), 1.18 (d, J = 5.8 Hz, 3H, Me- γ), 1.07 ppm (s, 3H, Me- β). This carboxylic acid (35.5 mg, 1.02×10^{-1} mmol) was dissolved in 0.3m HCl/AcOEt (1 mL). After stirring at room temperature for 1 h, the precipitates were filtered and washed with AcOEt to afford 50 (HCl salt, 25.3 mg, 87%) as yellow solids: ¹H NMR (CF₃CO₂D): $\delta = 8.69$ (s, 1H, thiazole H-5), 5.56 (brs, 1H, H- α), 4.34 (brq, J=6.2 Hz, 1H, H- γ), 1.53 (d, $J=6.2$ Hz, 3H, Me- γ), 1.23 ppm (s, 3H, Me- β) [Ref. [10d]: ¹H NMR $(CF₃CO₂H)$: 8.69 (s, 1H), 5.56 (s, 1H), 4.39 (m, 1H), 1.56 (d, 3H), 1.28 (s, 3H)]. Compound 50 (HCl salt) was applied to an ion-exchange chromatography (DOWEX 50W). Pyridine–acetic acid buffer (0.2m, pH 3.1) was used as eluent to afford 50 (AcOH salt) as yellow solids: α_{ID}^{28} -2.8 (c 1.00, 1 M AcOH) [Ref. [2c]: $[a]_D^{25}$ -4 (c 1, 1 M AcOH); Ref. [8a]: $[a]_D^{20}$

 -2.8 (c 1, AcOH); Ref. [10d]: $[\alpha]_D^{25}$ -4.4 (c 1, 1 M AcOH)]. The optical rotation of the enantiomer of 50, prepared from the enantiomer of 41, was $+4.4$ (c 1.00, 1 M AcOH, 28°C). In addition, the ¹H NMR spectrum of the diastereomer of 50, prepared from the diastereomer of 49, was different from that of 50.

TES ether 34: To a solution of 49 (300 mg, 8.01×10^{-1} mmol) in dry CH_2Cl_2 (8 mL) at 0 °C under Ar atmosphere were added 2,6-lutidine (0.560 mL, 4.81 mmol) and TESOTf (0.725 mL, 3.20 mmol). After stirring at room temperature for 1 h, the reaction mixture was quenched with H₂O (20 mL) and the mixture was extracted with CHCl₃ (20 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (20% AcOEt/hexane) to afford 34 (371 mg, 92%) as a colorless syrup: R_f = 0.70 (40% AcOEt/hexane); $[\alpha]_D^{24}$ -23.5 (c 1.00, CHCl₃); IR (neat): \tilde{v} = 3385, 2956, 2905, 2880, 1720, 1458, 1418, 1370, 1322, 1238, 1200, 1118, 1012, 960, 740, 724 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.11 (s, 1H, thiazole H-5), 4.39 (dq, $J=2.4$, 7.0 Hz, 2H, CO₂CH₂CH₃), 4.31 (s, 1H, H- α), 4.04 (q, $J=6.2$ Hz, 1H, H- γ), 1.38 (t, $J=7.0$ Hz, 3H, CO₂CH₂CH₃), 1.26 (s, 3H, Me- β), 1.18 (d, J=6.2 Hz, 3H, Me- γ), 0.92 (t, J=7.8 Hz, 9H, Si- (CH_2CH_3) ₃), 0.91 (t, J=7.8 Hz, 9H, Si(CH₂CH₃)₃), 0.68–0.46 ppm (m, 12H, Si(CH₂CH₃)₂×2); ¹³C NMR (CDCl₃); δ =175.93, 161.63, 146.30, 127.86, 80.34, 71.70, 61.06, 60.48, 19.28, 18.09, 14.24, 7.17, 6.83, 6.73, 5.08 ppm; HRMS (FAB): m/z $[M+H]^+$ calcd for $C_{23}H_{47}N_2O_4SSi_2$: 503.2802; found: 503.2795.

 β -Hydroxyamide pentapeptide 32: To a solution of 34 (199 mg, 3.96 \times 10^{-1} mmol), 33 (275 mg, 4.36×10^{-1} mmol), and HOAt (64.7 mg, 4.75 \times 10^{-1} mmol) in dry CH₂Cl₂ (4.0 mL) at 0°C under Ar atmosphere were added *iPr₂NEt* (0.166 mL, 9.53×10^{-1} mmol) and CIP (132 mg, 4.74 \times 10^{-1} mmol). After stirring at room temperature for 0.5 h, the reaction mixture was quenched with saturated aqueous $NaHCO₃$ (5 mL) and the mixture was extracted with AcOEt (10 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (30% AcOEt/hexane) to afford 32 (364 mg, 83% from 34) as a colorless foam: $R_f = 0.40$ (40% AcOEt/ hexane); $[\alpha]_D^{26}$ -15.5 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu}$ = 3685, 3620, 3420, 1686, 1602, 1518, 1478, 1420, 1044 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.09 (s, 1H, thiazole H-5), 7.60–7.48 (m, 2H, PhSe), 7.38 (d, J=9.2 Hz, 1H, CONH), 7.32-7.18 (m, 3H, PhSe), 7.14-6.74 (m, 2H, CONH \times 2), 5.39 (d, $J=9.2$ Hz, 1H, Ile H- α), 4.70–4.56 (m, 1H, phenylselenoamino acid Hα), 4.40–4.04 (m, 3H, Ser H-α and Me₃SiCH₂CH₂), 4.34 (q, $J=6.8$ Hz, 2H, CO₂CH₂CH₃), 4.02-3.80 (m, 3H, Ser H- $\beta \times 2$ and oxazolidine H-5), 3.80–3.52 (m, 3H, phenylselenoamino acid H-b, Ile H-g, and oxazolidine H-4), 1.63 (s, 3H, Me), 1.59 (s, 3H, Me), 1.46 (d, J=6.8 Hz, 3H, Me), 1.34 (t, $J=6.8$ Hz, 3H, $CO_2CH_2CH_3$), 1.29 (s, 3H, Me), 1.08 (d, $J=$ 6.0 Hz, 3H, Me), 0.96 (t, $J=9.0 \text{ Hz}$, 9H, Si(CH₂CH₃)₃), 0.84 (t, $J=$ 9.0 Hz, 9H, Si(CH₂CH₃)₃), 0.89–0.76 (m, 2H, Me₃SiC<u>H</u>₂CH₂), 0.68 (q, J= 9.0 Hz, 6H, Si(CH₂CH₃)₃), 0.58–0.44 (m, 6H, Si(CH₂CH₃)₃), 0.02 ppm (s, 9H, \underline{Me}_3 SiCH₂CH₂); ¹³C NMR (CDCl₃): $\delta = 170.01$, 169.38, 169.32, 168.12, 161.19, 153.50 (br), 145.56, 135.02, 129.18, 128.52, 128.09, 127.91, 95.09, 78.23, 74.10, 72.30, 67.41, 64.10, 62.71, 61.24, 60.88, 57.28, 56.36, 40.32, 27.40, 25.45, 19.20, 18.98, 18.47, 17.91, 17.67, 14.24, 7.14, 7.02, 6.81, 6.76, 5.05, 4.65, 1.61 ppm. Elemental analysis (%) calcd for $C_{49}H_{85}N_5O_{11}SSESi_3$: C 52.76, H 7.68, N 6.28; found: C 52.65, H 7.56, N 6.22.

 β -Hydroxythioamide pentapeptide 52: To a solution of 32 (516 mg, 4.63 \times 10^{-1} mmol) in dry CH₂Cl₂ (4.6 mL) at -78° C under Ar atmosphere was added DAST (0.0917 mL, 0.690 mmol) in dry CH_2Cl_2 (4.6 mL). After stirring at -78 °C for 5 min, the reaction mixture was quenched with saturated aqueous NaHCO₃ (10 mL) and extracted with AcOEt (15 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (20% AcOEt/hexane) to afford 51 (432 mg, 85%) as a colorless foam: $R_f = 0.50$ (25% AcOEt/hexane); ¹H NMR (CDCl₃): δ = 8.06 (s, 1H, thiazole H-5), 7.75 (d, J=9.4 Hz, 1H, CONH), 7.65–7.57 (m, 2H, PhSe), 7.34–7.22 (m, 3H, PhSe), 7.00–6.82 (m, 1H, CONH), 5.47 (d, J=9.4 Hz, 1H, Ile H-a), 5.04 (dd, $J=6.4$, 8.0 Hz, 1H, phenylselenoamino acid H- α), 4.70 (dd, $J=$ 8.2, 9.2 Hz, 1H, Ser H- α), 4.52-4.30 (m, 2H, Ser H- $\beta \times 2$), 4.36 (q, J= 7.0 Hz, 2H, $CO_2CH_2CH_3$), 4.24–4.02 (m, 3H, Me₃SiCH₂CH₂ and oxazoli-

dine H-5), 3.84–3.64 (m, 3H, oxazolidine H-4, phenylselenoamino acid H- β , and Ile H- γ), 1.62 (s, 3H, Me), 1.59 (s, 3H, Me), 1.52 (d, J=6.8 Hz, 3H, Me), 1.35 (t, $J=7.0$ Hz, 3H, CO₂CH₂CH₃), 1.34 (s, 3H, Me), 1.14 (d, $J=5.8$ Hz, 3H, Me), 1.11 (d, $J=6.0$ Hz, 3H, Me), 0.96 (t, $J=7.2$ Hz, 9H, $Si(CH_2CH_3)$ ₃), 0.91 (t, J = 7.2 Hz, 9H, Si(CH₂CH₃)₃), 1.04–0.80 (m, 2H, $Me₃SiCH₂CH₂$, 0.66 (q, J = 7.2 Hz, 6H, Si(CH₂CH₃)₃), 0.59 (q, J = 7.2 Hz, 6H, Si(CH_2CH_3)₃), 0.02 ppm (s, 9H, $Me_3SiCH_2CH_2$). A solution of 51 $(432 \text{ mg}, 3.94 \times 10^{-1} \text{ mmol})$ in MeOH (2.0 mL) and NEt₃ (2.0 mL) was saturated with H2S and stirred at room temperature for 10 h. Ar was bubbled through the reaction mixture for 15 min and the mixture was evaporated. The residue was chromatographed on silica gel (20% AcOEt/ hexane) to afford 52 (401 mg, 90%) as a colorless foam: $R_f = 0.40$ (30%) AcOEt/hexane); $[\alpha]_D^{27}$ +21.1 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 3685, 3620$, 3418, 1682, 1518, 1475, 1420, 1340, 1046 cm⁻¹; ¹H NMR (CDCl₃) δ = 8.60– 8.42 (m, 1H, CSNH), 8.11 (s, 1H, thiazole H-5), 7.61–7.48 (m, 2H, PhSe), 7.37–7.20 (m, 4H, CONH, PhSe), 7.05 (d, J=7.6 Hz, 1H, CONH), 5.42 (d, $J=9.2$ Hz, 1H, Ile H- α), 4.88–4.78 (m, 1H, phenylselenoamino acid H- α), 4.76–4.65 (m, 1H, Ser H- α), 4.37 (q, J=7.0 Hz, 2H, CO₂CH₂CH₃), 4.30-4.00 (m, 5H, Me₃SiCH₂CH₂, Ser H- β , oxazolidine H-5, and phenylselenoamino acid H-b), 4.00–3.82 (m, 2H, oxazolidine H-4 and Ser H- β), 3.66 (q, J=6.2 Hz, 1H, Ile H- γ), 1.65 (s, 3H, Me), 1.61 (s, 3H, Me), 1.46 (d, J=6.8 Hz, 3H, Me), 1.41 (d, J=6.2 Hz, 3H, Me), 1.36 (t, $J=7.0$ Hz, 3H, $CO_2CH_2CH_3$), 1.28 (s, 3H, Me), 1.09 (d, $J=6.2$ Hz, 3H, Me), 1.06–0.85 (m, 2H, Me₃SiCH₂CH₂), 0.98 (t, $J=7.8$ Hz, 9H, Si- $(CH_2C\underline{H}_3)$ ₃), 0.86 (t, J = 7.8 Hz, 9 H, Si(CH₂C_{H₃)₃), 0.71 (q, J = 6.8 Hz,} 6H, Si(CH₂CH₃)₃), 0.62–0.44 (m, 6H, Si(C<u>H₂</u>CH₃)₃), 0.20 ppm (s, 9H, $Me_3SiCH_2CH_2$); ¹³C NMR (CDCl₃): $\delta = 201.39, 168.87, 168.71, 168.12,$ 161.14, 154.50–151.50 (br), 145.53, 134.43, 129.24, 128.55, 128.18, 127.99, 95.06, 78.10, 74.14, 72.38, 67.56, 64.15, 62.82, 62.01, 61.77, 61.24, 60.99, 42.82, 27.42 (br), 25.31 (br), 19.29, 17.91, 17.61, 14.23, 7.14, 6.84, 6.74, 5.10, -1.63 ppm. Elemental analysis (%) calcd for $C_{49}H_{85}N_5O_{10}S_2SeSis3$: C 52.01, H 7.57, N 6.19; found: C 51.87, H 7.30, N 6.11.

Thiazoline 54: To a solution of 52 (33.1 mg, 2.93×10^{-2} mmol) in dry CH_2Cl_2 (0.3 mL) at $-78\degree$ C under Ar atmosphere was added DAST (0.0043 mL, 3.3×10^{-2} mmol) in dry CH₂Cl₂ (0.3 mL). After stirring at -78 °C for 5 min, the reaction mixture was quenched with saturated aqueous NaHCO₃ (2 mL) and the mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated to afford 53. To a solution of 53 in dry CH_2Cl_2 (0.15 mL) and TFE (0.15 mL) at 0°C under Ar atmosphere was added 4.89 M TBHP in CH₂Cl₂ (0.0598 mL, 0.292 mmol). After stirring at room temperature for 2 h, the reaction was quenched with saturated aqueous $Na₂S₂O₃$ (1.5 mL) and the mixture was extracted with AcOEt $(5 \text{ mL} \times 3)$. The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (25% AcOEt/hexane) to afford 54 (16.1 mg, 57% from 52) as a colorless foam: $R_{\rm f} = 0.60$ (30% AcOEt/hexane); $\left[\alpha\right]_{\rm D}^{26}$ -22.0 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 3685, 3620, 3405, 1706, 1520, 1478, 1420, 1338,$ 1118, 1046, 498 cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.09$ (s, 1H, thiazole H-5), 7.73 (br s, 1H, CONH), 7.55 (d, J=8.6 Hz, 1H, CONH), 6.48 (q, J= 7.0 Hz, 1 H, Δ Abu H- β), 5.45 (d, $J=8.6$ Hz, 1 H, Ile H- α), 5.02 (dd, $J=$ 9.2, 11.0 Hz, 1H, Ser H- α), 4.35 (q, J = 7.0 Hz, 2H, CO₂CH₂CH₃), 4.42– 4.24 (m, 1H, oxazolidine H-5), 4.24-4.02 (m, 2H, Me₃SiCH₂CH₂), 3.96 (d, $J=7.4$ Hz, 1H, oxazolidine H-4) 3.86-3.66 (m, 2H, Ser H- β , Ile H- γ), 3.57 (dd, $J=9.2$, 11.0 Hz, 1H, Ser H- β), 1.82 (d, $J=7.0$ Hz, 3H, Δ Abu Me- β), 1.67 (s, 3H, Me), 1.64 (s, 3H, Me), 1.46 (d, J = 5.6 Hz, 3H, Me), 1.36 (t, $J=7.0$ Hz, 3H, $CO_2CH_2CH_3$), 1.35 (s, 3H, Me), 1.12 (d, $J=$ 6.0 Hz, 3H, Me), 1.06–0.87 (m, 2H, Me₃SiCH₂CH₂), 0.94 (t, $J=7.6$ Hz, 9H, Si(CH₂CH₃)₃), 0.87 (t, J=7.6 Hz, 9H, Si(CH₂CH₃)₃), 0.72-0.42 (m, 12H, Si(CH₂CH₃)₃×2), 0.20 ppm (s, 9H, Me₃SiCH₂CH₂); ¹³C NMR (CDCl₃): $\delta = 171.11$, 170.86, 168.74, 167.34, 161.37, 152.80 (br), 146.01, 131.99, 128.16, 95.29, 79.47, 79.14, 74.65 (br), 72.02, 67.76, 63.79, 61.11, 59.37, 36.16, 27.15 (br), 25.20, 19.26, 18.95, 17.95, 17.78, 15.08, 14.33, 7.20, 6.91, 6.76, 5.08, -1.60 ppm; HRMS (FAB): m/z [$M+H$]⁺ calcd for $C_{43}H_{78}N_5O_9S_2Si_3$: 956.4519; found: 956.4548.

Segment B (3) : To a solution of 52 (30.0 mg, 2.65×10^{-2} mmol) in EtOH (0.180 mL) and 1,4-dioxane (0.090 mL) at 0° C was added 1M aqueous NaOH (0.080 mL). After stirring at room temperature for 5 h, the reaction mixture was acidified with 1 μ aqueous HCl to pH 3 at 0°C and ex-

tracted with AcOEt $(1 mL \times 3)$. The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated to afford 3 as a colorless foam, which was used for the next step without purification: $R_f=0.10$ (30% AcOEt/hexane); $[\alpha]_D^{24}$ + 22.6 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu} = 3680$, 3620, 3418, 1748, 1682, 1520, 1478, 1420, 1338, 1044, 498 cm⁻¹; ¹H NMR $(CDCI_3)$; $\delta = 9.14-8.90$ (brs. 1H, CSNH), 8.21 (s, 1H, thiazole H-5), 7.62–7.48 (m, 2H, PhSe), 7.48–7.35 (m, 1H, CONH), 7.34–7.18 (m, 3H, PhSe), 6.95–6.66 (m, 1H, CONH), 5.52–5.20 (m, 1H, Ile H- α), 5.10–4.66 (m, 2H, Ser H- α and phenylselenoamino acid H- α), 4.43–4.17 (m, 4H, $Me₃SiCH₂CH₂$, oxazolidine H-5, and phenylselenoamino acid H- β), 4.12 (br dd, 1H, Ser H- β), 4.01 (br dd, 1H, Ser H- β) 3.93 (d, J=7.6 Hz, 1H, oxazolidine H-4), 3.73 (br q, 1H, Ile H- γ), 1.66 (s, 6H, Me), 1.42 (d, J= 6.0 Hz, 3H, Me), 1.38–1.18 (m, 6H, Me \times 2), 1.11 (d, J = 5.8 Hz, 3H, Me), 1.05–0.95 (m, 2H, Me₃SiCH₂CH₂), 0.98 (t, J = 7.2 Hz, 9H, Si(CH₂CH₃)₃), 0.84 (t, $J=7.2$ Hz, 9H, Si(CH₂CH₃)₃), 0.72 (q, $J=7.8$ Hz, 6H, Si- (CH_2CH_3) , 0.52 (q, J=7.8 Hz, 6H, Si (CH_2CH_3) ₃), 0.20 ppm (s, 9H, $Me₃SiCH₂CH₂$).

Aminoalcohol 55: To a solution of 52 (32.0 mg, 2.83×10^{-2} mmol) in $CH₃NO₂$ (0.28 mL) at 0°C under Ar atmosphere was added 1.0 m ZnCl₂ in ether (0.42 mL, 4.2×10^{-1} mmol). After stirring at room temperature for 15 h, the reaction mixture was quenched with saturated aqueous NaHCO₃ (1 mL) and the mixture was extracted with AcOEt (3 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (5–10% MeOH/CHCl₃) to afford 55 (15.1 mg, 58%) as a colorless foam: R_f = 0.50 (8% MeOH/CHCl₃); $[\alpha]_D^{26}$ +41.1 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu} = 3682$, 3620, 3420, 1722, 1676, 1518, 1478, 1420, 1044, 498 cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.90 - 8.72$ (m, 1H, CSNH), 8.11 (s, 1H, thiazole H-5), 8.11– 8.04 (m, 1H, CONH), 7.62–7.54 (m, 2H, PhSe), 7.38–7.20 (m, 4H, CONH, PhSe), 5.43 (d, $J=9.2$ Hz, 1H, Ile H- α), 4.98–4.87 (m, 1H, phenylselenoamino acid H- α), 4.81–4.72 (m, 1H, Ser H- α), 4.38 (q, J= 7.0 Hz, 2 H, CO₂CH₂CH₃), 4.27 (dq, $J = 3.2$, 6.4 Hz, 1 H, oxazolidine H-5), 4.18–4.04 (m, 2H, Ser H- β and phenylselenoamino acid H- β), 3.94 (dd, $J=3.6$, 12.4 Hz, 1 H, Ser H- β), 3.71 (q, $J=6.4$ Hz, 1 H, Ile H- γ), 3.37 (br s, 1H, oxazolidine H-4), 1.41 (d, $J=7.0$ Hz, 3H, Me), 1.37 (t, $J=7.0$ Hz, 3H, CO₂CH₂CH₃), 1.29 (s, 3H, Me), 1.21 (d, J = 6.4 Hz, 3H, Me), 1.08 (d, $J=6.4$ Hz, 3H, Me), 0.97 (t, $J=7.2$ Hz, 9H, Si(CH₂CH₃)₃), 0.87 (t, $J=$ 7.2 Hz, 9H, Si(CH₂CH₃)₃), 0.69 (q, J=7.2 Hz, 6H, Si(C<u>H₂CH₃)₃)</u>, 0.59– 0.48 ppm (m, 6H, Si(CH₂CH₃)₃); ¹³C NMR (CDCl₃): δ = 201.77, 173.79, 169.35, 168.48, 161.21, 145.56, 134.71, 129.23, 128.59, 128.44, 128.03, 78.40, 72.26, 68.03, 63.07, 61.73, 61.57, 61.36, 60.63, 59.63, 42.75, 29.67, 19.54, 18.69, 17.80, 17.39, 14.28, 7.20, 6.89, 6.78, 5.11 ppm; HRMS (FAB): m/z $[M+H]^+$ calcd for $C_{40}H_{70}N_5O_8S_2SeSi_2$: 948.3375; found: 948.3369.

Epoxy alcohols 67 and 68 from 63: To a solution of 63 (100 mg, $3.20 \times$ 10^{-1} mmol) in dry toluene (3.2 mL) at -78° C under Ar atmosphere were added HMPA (0.0613 mL, 3.52×10^{-1} mmol) and 3m MeMgBr in Et₂O (0.117 mL, 3.51×10^{-1} mmol). After stirring at -78 °C for 0.5 h, the reaction mixture was quenched with saturated aqueous NH4Cl (5 mL) and the mixture was extracted with AcOEt ($5 \text{ mL} \times 3$). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (30% AcOEt/hexane) to afford 67 (50.0 mg, 48%) as a colorless foam, 68 (15.8 mg, 15%) as a colorless foam, and the recovered 63 (20.0 mg, 20%) as a colorless foam. 67: R_f = 0.39 (70% AcOEt/hexane); m.p. 125–126 °C (not recrystallized); $\lbrack a \rbrack_{D}^{28}$ -22.3 (91% ee) (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 3620, 1730, 1440, 1422,$ 1309, 1125, 876 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.40 (s, 1H, H-3), 5.39 (dq, $J=3.2, 6.4$ Hz, 1H, CH(CH₃)OH), 5.34 (ddd, $J=1.8$, 1.8, 5.4 Hz, 1H, H-5), 4.40 (d, $J=3.4$ Hz, 1H, H-8), 4.02 (s, 3H, CO₂Me), 3.98-3.93 (m, 1H, H-7), 3.12 (ddd, $J=1.8$, 1.8, 16.8 Hz, 1H, H-6), 2.51 (ddd, $J=0.9$, 5.4, 16.8 Hz, 1H, H-6), 2.21 (d, J=3.2 Hz, 1H, CH(CH₃)O<u>H</u>), 1.51 ppm (d, $J=6.4$ Hz, 3H, CH(CH₃)OH); ¹³C NMR (CDCl₃): $\delta=165.07$, 154.68, 152.54, 147.91, 130.48, 122.55, 64.19, 54.04, 53.10, 34.92, 30.38, 24.61 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C₁₃H₁₅NO₄Br: 328.0184; found: 328.0187. 68: $R_f = 0.24$ (70% AcOEt/hexane); m.p. 128-129 °C (not recrystallized); $[a]_D^{29} + 21.0$ (91 % ee) (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu} = 3620$, 1728, 1440, 1423, 1310, 1128, 879 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.29 (s, 1H, H-3), 5.66 (ddd, J=1.5, 1.5, 5.2 Hz, 1H, H-5), 5.30 (dq, J=4.2, 6.4 Hz, 1H, CH(CH3)OH), 4.41 (d, J=1.5, 1.5, 3.8 Hz, 1H, H-8), 4.02 (s, 3H, CO₂Me), 3.99–3.94 (m, 1H, H-7), 3.13 (ddd, $J=1.5$, 1.5, 16.8 Hz, 1H, H-

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6), 2.53 (ddd, $J=0.8$, 5.4, 16.8 Hz, 1H, H-6), 1.94 (d, $J=4.2$ Hz, 1H, CH-(CH₃)O<u>H</u>), 1.66 ppm (d, $J=6.4$ Hz, 3H, CH(CH₃)OH); ¹³C NMR (CDCl₃): δ = 165.14, 153.13, 147.75, 131.78, 122.85, 64.10, 54.23, 53.38, 53.15, 34.81, 30.83, 23.69 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C13H15NO4Br: 328.0184; found: 328.0181.

Methoxyquinoline 69: To a solution of 67 (24.5 mg, 7.47×10^{-2} mmol) in dry THF (0.75 mL) at 0°C under Ar atmosphere was added DBU (0.0335 mL, 2.24×10^{-1} mmol). The reaction mixture was stirred at room temperature for 1 h and then 1m aqueous HCl (0.75 mL) was added at 0° C. After stirring at room temperature for 1 h, to the reaction mixture was added H₂O (1 mL), and the mixture was extracted with AcOEt $(2 mL \times 3)$. The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated to afford the crude 8-quinolinol (18.2 mg). The crude 8-quinolinol was dissolved in MeOH (0.6 mL), followed by addition of CH_2N_2 in Et₂O. After stirring at room temperature for 1 h, the reaction mixture was quenched with saturated aqueous $NaHCO₃$ (2 mL) and the mixture was extracted with AcOEt ($2 \text{ mL} \times 3$). The combined extracts were dried over Na_2SO_4 , filtered through celite, and evaporated. The residue was chromatographed on silica gel (15% acetone/hexane) to afford 69 (15.9 mg, 83%) as colorless solids: R_f = 0.46 (30% acetone/ CHCl₃); $[\alpha]_{D}^{28}$ -76.3 (c 1.00, EtOH); UV (EtOH) λ_{max} nm (log ε): 343 (3.15), 303 (3.23), 254 (4.18); m.p. 170–171 8C (not recrystallized); IR (nujol): $\tilde{v} = 3400, 1735$ cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.38$ (s, 1H, H-3), 7.58 (d, $J=3.4$ Hz, 1H, H-5), 7.55 (d, $J=4.8$ Hz, 1H, H-7), 7.05 (dd, $J=3.4$, 4.8 Hz, 1H, H-6), 5.61 (q, J=6.6 Hz, 1H, CH(CH3)OH), 4.08 (s, 3H, CO₂Me), 4.04 (s, 3H, OMe), 2.55 (brs, CH(CH₃)O<u>H</u>), 1.65 ppm (d, $J=$ 6.6 Hz, 3H, CH(C<u>H</u>₃)OH); ¹H NMR (CD₃CO₂D): δ = 8.37 (s, 1H, H-3), 7.60 (m, 2H, aromatic H), 7.16 (d, J=7.2 Hz, 1H, aromatic H), 5.70 (q, $J=6.5$ Hz, 1H, CH(CH₃)OH), 4.01 (s, 3H, CO₂Me), 3.98 (s, 3H, OMe), 1.61 ppm (d, J=6.5 Hz, 3H, CH(C<u>H</u>₃)OH); ¹³C NMR(CDCl₃): δ = 166.06, 156.44, 152.80, 146.52, 139.63, 129.08, 127.58, 117.22, 114.49, 107.66, 66.46, 56.14, 53.05, 24.35 ppm; HRMS (FAB): m/z [$M+H$]⁺ calcd for C₁₄H₁₆NO₄: 262.1079; found: 262.1081 [Ref. [8a]: $[\alpha]_D^{20}$ –79 (c 1.00, EtOH); m.p. darkened at 161–168 °C, melted at 174 °C; IR (nujol): \tilde{v} = 3300, 1735 cm⁻¹; UV $\lambda_{\text{max}}^{\text{alc}}$ nm $(E_{1\text{cm}}^{1\%})$: 347 (128), 317, 307, 254 (1553); ¹H NMR (CD CO D); $\lambda = 8.37$ (aromatic H) 7.58 (side chain qH) 7.16 ¹H NMR (CD₃CO₂D): δ = 8.37 (aromatic H), 7.58 (side chain α H), 7.16, 4.00 (OMe), 1.60 ppm (d, 3H). Elemental analysis (%) calcd for $C_{14}H_{15}NO_4$: C 64.36, H 5.79, N 5.36, O 24.49; found: C 63.72, H 6.00, N 5.48, O 24.05; Ref. [2c]: α_{D}^{20} –78 (c 1.6, EtOH); m.p. about 161–168 °C to 175–177°C; IR: $\tilde{\nu} = 3400, 1750 \text{ cm}^{-1}$; UV $\lambda_{\text{max}}^{\text{alc}}$ nm $(E_{1 \text{cm}}^{1\%})$: 347 (115), 254 (1500); ¹H NMR (CD₃CO₂D): $\delta = 8.24$ (s, pyridine H-3), 7.50 (d, 2H), 7.17 (m, 1H), 5.63 (d, 1H, side-chain a-carbon proton), 3.96 (s, 6H, OMe), 1.59 ppm (d, $J=6.5$ Hz, 3H). Elemental analysis (%) calcd for $C_{14}H_{15}NO_4$: C 64.36, H 5.79, N 5.36; found: C 64.50, H 6.09, N 5.58].

Dihydroquinoline 59: To a solution of 61 (9.50 g, 49.7 mmol) in acetaldehyde (350 mL) and H₂O (250 mL) at 0° C were slowly added TFA (3.69 mL, 49.7 mmol), $FeSO_4$ -7 H_2O (1.38 g, 4.96 mmol), and 31% aqueous H_2O_2 (9.74 mL, 97.7 mmol). After stirring at room temperature for 4 h, the reaction mixture was quenched with saturated aqueous $NaHCO₃$ (500 mL), saturated aqueous $\text{Na}_2\text{S}_2\text{O}_3$ (250 mL), and H₂O (250 mL). The mixture was extracted with AcOEt $(1 L \times 3)$. The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (5% acetone/CHCl₃) to afford methyl 4-acetyl-5,6,7,8-tetrahydroquinoline-2-carboxylate 60 (9.73 g, 84%) as colorless solids: $R_f = 0.62$ (50% AcOEt/hexane); ¹H NMR (CDCl₃): $\delta =$ 8.05 (s, 1H, H-3), 4.02 (s, 3H, CO₂Me), 3.10 (t, $J=6.0$ Hz, 2H, H-8), 2.97 $(t, J=6.0 \text{ Hz}, 2\text{ H}, \text{ H-5}), 2.61 \text{ (s, 3H, CH₃C(O)), 1.91-1.82 ppm (m, 4H,$ H-6 and H-7). To a solution of 60 (11.3 g, 48.4 mmol) in CH₂Cl₂ (162 mL) at 0°C was slowly added 65% MCPBA (25.7 g, 96.8 mmol). After stirring at room temperature for 7 h, saturated aqueous NaHCO₃ (150 mL), saturated aqueous $Na_2S_2O_3$ (100 mL), and H_2O (100 mL) were slowly added and the resulting mixture was extracted with CHCl₃ (250 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (40% acetone/hexane) to afford 70 (10.6 g, 88%) as colorless solids. To a solution of 70 (7.00 g, 28.1 mmol) in dry CH₂Cl₂ (280 mL) at 0 °C under Ar atmosphere was added dropwise Tf_2O (5.60 mL, 33.3 mmol) over 10 min. After stirring at 0° C for 10 min, a solution of NEt₃ (19.6 mL, 141 mmol) in dry $CH₂Cl₂$ (280 mL) was added dropwise over 1 h. The reaction mixture was stirred at room temperature for 5 h, quenched with $H₂O$ (450 mL), and extracted with CHCl₃ (1 L \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel $(40:60:2$ acetone/hexane/NEt₃) to afford 59 (6.36 g, 98%) as a colorless syrup: $R_f = 0.63$ (70% AcOEt/hexane); m.p. 118–120 °C (not recrystallized); IR (CHCl₃): $\tilde{v} = 1720, 1695, 1450, 1425,$ 1315, 1240, 1155, 980, 895, 785 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.08 (s, 1H, H-3), 6.82 (dt, J=1.6, 10.0 Hz, 1H, H-8), 6.49 (dt, J=4.5, 10.0 Hz, 1H, H-7), 4.02 (s, $3H$, CO₂Me), 3.12 (t, $J=8.0$ Hz, $2H$, H-5), 2.64 (s, $3H$, CH₃C(O)), 2.45–2.32 ppm (m, 2H, H-6); ¹³C NMR (CDCl₃): δ = 200.63, 165.46, 155.26, 145.51, 144.11, 136.06, 132.81, 129.20, 121.12, 52.98, 29.73, 23.93, 22.12 ppm; HRMS (EI): m/z [M]⁺ calcd for C₁₃H₁₃NO₃: 231.0895; found: 231.0889.

Epoxytetrahydroquinoline 79: To a solution of 59 (13.9 mg, 0.0601 mmol) and 97% 4-phenylpyridine N-oxide (5.1 mg, 0.030 mmol) in CH₃CN (0.12 mL) at -10 °C were added (R,R)-Mn-salen catalyst 81 (2.9 mg, 0.0046 mmol) and iodosobenzene (26.4 mg, 0.120 mmol). After stirring at -10 °C for 15 h, the reaction mixture was filtered through celite and evaporated. $H₂O$ (2 mL) was added and the mixture was extracted with AcOEt (2 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (20% acetone/hexane) to afford epoxide 79 (10.9 mg, 73%, 82% ee) as colorless solids. The enantiomeric exess of 79 was determined by chiral HPLC analysis (Daicel Chiralcel OD column, 4.6×250 mm, 90:10 hexane–IPA; 1 mLmin^{-1} , 254 nm , $t = 20.6 \text{ min}$; enantiomer of 79, $t=30.8$ min). **79**: $R_f=0.40$ (30% acetone/hexane); m.p. 90–93 °C (not recrystallized); $[\alpha]_D^{27}$ +58.8 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu} = 3010, 2960,$ 1715, 1685, 1550, 1440, 1410, 1300, 1245, 1155, 1075, 1005, 820, 780 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.20 (s, 1H, H-3), 4.29 (d, J = 4.0 Hz, 1H, H-8), 4.05 (s, 3H, CO₂Me), 3.85 (m, 1H, H-7), 3.06 (dddd, $J=1.4$, 1.4, 5.4, 17.2 Hz, 1H, H-5), 2,76 (ddd, J=6.8, 13.2, 17.2 Hz, 1H, H-5), 2.61 (s, 3H, CH3C(O)), 2.57–2.43 (m, 1H, H-6), 1.76 ppm (ddd, J=5.4, 13.2, 14.1 Hz, 1H, H-6); ¹³C NMR (CDCl₃): δ = 200.42, 165.00, 155.45, 145.70, 145.51, 133.74, 122.40, 55.10, 53.81, 53.15, 30.01, 20.88, 20.30 ppm; HRMS (EI): m/z [M]⁺ calcd for C₁₃H₁₃NO₄: 247.0845; found: 247.0846 [80: $R_f = 0.56$ (30% acetone/hexane); ¹H NMR (CDCl₃): $\delta = 8.55$ (dd, $J = 1.0$, 8.8 Hz, 1H, H-8), 8.45 (s, 1H, H-3), 8.36 (brd, $J=8.4$, 1H, H-5), 7.86 (ddd, $J=$ 0.8, 7.6, 8.8 Hz, 1 H, H-7), 7.76 (ddd, $J=1.0$, 7.6, 8.4 Hz, 1 H, H-6), 4.13 (s, 3H, $CO₂Me$), 2.82 ppm (s, 3H, $CH₃C(O)$)].

Epoxybromide 58 and its epimer 82: To a solution of 79 (4.80 g, 19.4 mmol) in CCl_4 (200 mL) were added NBS (3.83 g, 21.5 mmol) and AIBN (321 mg, 1.95 mmol). The reaction mixture was stirred and irradiated with a 140-W sun lamp at 60°C for 5 h. H₂O (200 mL), saturated aqueous $Na₂S₂O₃$ (100 mL), and saturated aqueous NaHCO₃ (100 mL) were added and the mixture was extracted with CHCl₃ (500 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (50% AcOEt/hexane) to afford 58 (4.28 g, 67%) as colorless solids and 82 (685 mg, 11%) as colorless solids. **58**: $R_f = 0.45$ (40% AcOEt/CHCl₃); m.p. 159–161 °C (not recrystallized); $[\alpha]_{D}^{28}$ –92.7 (c 1.00, CHCl₃); IR $(CHCl₃)$: $\tilde{v} = 3085$, 3005, 2955, 1720, 1560, 1420, 1320, 1250, 1150, 1130, 780 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.19 (s, 1H, H-3), 6.04 (ddd, J = 1.8, 1.8, 5.8, Hz, 1H, H-5), 4.40 (d, $J=3.8$ Hz, 1H, H-8), 4.07 (s, 3H, CO₂Me), 4.01–3.95 (m, 1H, H-7), 3.07 (ddd, J=1.8, 1.8, 17.2 Hz, 1H, H-6), 2.68 (s, 3H, CH3C(O)), 2.57 ppm (br dd, J=5.8, 17.2 Hz, 1H, H-6); 13C NMR (CDCl₃): δ = 200.70, 164.46, 154.18, 147.63, 145.72, 132.12, 122.49, 54.45, 53.37, 53.04, 33.63, 30.00, 29.78 ppm; HRMS (EI): m/z [M]⁺ calcd for $C_{13}H_{12}NO_4^{79}Br: 324.9950$; found: 324.9960. **82**: $R_f = 0.61(40\%$ AcOEt/ CHCl₃); ¹H NMR (CDCl₃): δ = 8.22 (s, 1H, H-3), 5.86 (dd, J = 4.8, 5.2 Hz, 1H, H-5), 4.23 (d, $J=3.8$ Hz, 1H, H-8), 4.07 (s, 3H, CO₂Me), 3.94–3.86 $(m, 1H, H-7)$, 2.94 (ddd, $J=3.5, 5.2, 16.0$ Hz, 1H, H-6), 2.71 (ddd, $J=1.8$, 4.8, 16.0 Hz, 1H, H-6), 2.68 ppm (s, 3H, CH₃C(O)).

Epoxy alcohols 67 and 68 from 58 : To a solution of 58 $(4.60 g,$ 14.1 mmol) in MeOH (141 mL) at -78° C under Ar atmosphere was added NaBH₄ (2.40 g, 63.4 mmol). After stirring at -78 °C for 19 h, the reaction mixture was quenched with saturated aqueous NH4Cl (50 mL) and H₂O (50 mL). The mixture was extracted with AcOEt (100 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite,

and evaporated. The residue was chromatographed on silica gel (60% AcOEt/hexane) to afford alcohol 67 (4.38 g, 95%) as a colorless foam and 68 (170 mg, 4%) as a colorless foam.

TBS ether 85: To a solution of 67 (20.6 mg, 6.28×10^{-2} mmol) in dry CH_2Cl_2 (0.63 mL) at 0 °C under Ar atmosphere were added 2,6-lutidine $(0.0146 \text{ mL}, \quad 1.25 \times 10^{-1} \text{ mmol})$ and TBSOTf $(0.0173 \text{ mL}, \quad 7.53 \times 10^{-1} \text{ mmol})$ 10^{-2} mmol). After stirring at 0° C for 0.5 h, the reaction mixture was quenched with H_2O (2 mL) and saturated aqueous NaHCO₃ (0.50 mL). The mixture was extracted with $ACOE$ (1 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (40% AcOEt/hexane) to afford the silylated bromide (22.7 mg, 82%) as a colorless syrup: R_f = 0.65 (70% AcOEt/hexane); $\left[\alpha\right]_D^{29}$ -35.3 (91% ee) (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 3622, 1730, 1440, 1424, 1315, 1300, 1130, 875$ cm⁻¹; ¹H NMR (CDCl₃): $\delta = 8.37$ (s, 1H, H-3), 5.31–5.22 (m, 2H, H-5 and CH- $(CH₃)$ OTBS), 4.37 (d, J = 3.9 Hz, 1H, H-8), 4.00 (s, 3H, CO₂Me), 3.94– 3.90 (m, 1H, H-7), 3.08 (br d, $J=16.8$ Hz, 1H, H-6), 2.46 (br dd, $J=5.2$, 16.8 Hz, 1 H, H-6), 1.39 (d, $J=6.3$ Hz, 3 H, CH(CH₃)OTBS) 0.91 (s, 9 H, $\sinh(2tBu)$, 0.09 (s, 3H, $\sinh(2tBu)$, -0.05 ppm (s, 3H, $\sinh(2tBu)$; ¹³C NMR (CDCl₃): δ = 165.26, 155.62, 152.37, 147.84, 129.95, 122.77, 65.38, 53.91, 53.22, 34.85, 30.48, 26.58, 25.70, 17.97, 4.78, 4.93 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C₁₉H₂₉NO₄Si⁸¹Br: 444.1029; found: 444.1032. To a solution of this sample (1.17 g, 2.64 mmol) in dry THF (26 mL) at 0° C under Ar atmosphere was added DBU (1.38 mL, 9.23 mmol). After stirring at room temperature for 1 h, the reaction mixture was quenched with $H₂O$ (20 mL) and the mixture was extracted with AcOEt (30 mL \times 3). The combined organic layers were washed with brine, dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (40% AcOEt/hexane) to afford 85 (909 mg, 95%) as a colorless foam: $R_f = 0.35$ (30% AcOEt/ hexane); ¹H NMR (CDCl₃): $\delta = 8.31$ (s, 1H, H-3), 7.08 (dd, J=1.4, 10.0 Hz, 1 H, H-5), 6.67 (dd, $J=3.4$, 10.0 Hz, 1 H, H-6), 5.17 (q, $J=$ 6.5 Hz, 1H, CH(CH₃)OTBS), 4.82 (d, $J=3.8$ Hz, 1H, H-8), 4.16 (ddd, $J=$ 1.4, 3.4, 3.8 Hz, 1 H, H-7), 4.02 (s, 3 H, CO₂Me), 1.40 (d, $J=6.5$ Hz, 3 H, $CH(CH₃)OTBS$), 0.87 (s, 9H, SiMe_{2t}Bu), 0.04 (s, 3H, SiMe_{2t}Bu), -0.08 ppm (s, 3H, SiMe_{2t}Bu); ¹³C NMR (CDCl₃): $\delta = 165.43, 153.05,$ 152.07, 146.33, 129.24, 126.18, 125.12, 122.56, 67.53, 58.51, 53.16, 52.96, 25.97, 25.63, 18.02, 4.93, 5.01 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C19H28NO4Si: 362.1788; found: 362.1787.

tert-Butyl ester 56: To a solution of 85 (12.8 mg, 3.54×10^{-2} mmol) in dry Et₂O (0.35 mL) at 0° C under Ar atmosphere was added 90% TMSOK (5.1 mg, 3.6×10^{-2} mmol). After stirring at 0 °C for 0.5 h, the reaction mixture was quenched with saturated aqueous NH_2Cl (2 mL) and the mixture was extracted with AcOEt $(2 mL \times 3)$. The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated to afford 86 (12.4 mg, quantitative yield) as colorless solids: ¹H NMR (CDCl₃): δ = 8.41 (s, 1H, H-3), 7.15 (dd, J=1.8, 9.8 Hz, 1H, H-5), 6.74 (dd, J=3.8, 9.8 Hz, 1 H, H-6), 5.21 (q, $J=6.0$ Hz, 1 H, CH(CH₃)OTBS), 4.73 (d, $J=$ 3.8 Hz, 1H, H-8), 4.23 (ddd, J=1.8, 3.8, 3.8 Hz, 1H, H-7), 1.43 (d, J= 6.0 Hz, 3H, CH(CH₃)OTBS) 0.89 (s, 9H, SiMe₂tBu), 0.08(s, 3H, Si- \underline{Me}_2tBu , -0.04 ppm (s, 3H, Si \underline{Me}_2tBu). To a solution of 86 (12.4 mg, 3.54×10^{-2} mmol) in tBuOH (0.35 mL) under Ar atmosphere were added DMAP $(1.3 \text{ mg}, 1.1 \times 10^{-2} \text{ mmol})$ and Boc₂O $(0.0163 \text{ mL}, 7.09 \times$ 10^{-2} mmol). After stirring at room temperature for 3 h, the reaction mixture was diluted with toluene (2 mL) and evaporated. The residue was chromatographed on silica gel (30% AcOEt/hexane) to afford 56 (12.3 mg, 86% from 85) as a colorless foam: $R_f = 0.65$ (50% AcOEt/ hexane); $[\alpha]_D^{26}$ -3.4 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu} = 3620, 2975, 1720,$ 1370, 1310, 1150, 1140, 1090, 1050, 880 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.23 (s, 1H, H-3), 7.01 (dd, J=1.8, 9.9 Hz, 1H, H-5), 6.65 (dd, J=3.6, 9.9 Hz, 1H, H-6), 5.18 (q, $J=6.3$ Hz, 1H, CH(CH₃)OTBS), 4.84 (d, $J=3.6$ Hz, 1H, H-8), 4.16 (ddd, $J=1.8$, 3.6, 3.6 Hz, 1H, H-7), 1.64 (s, 1H, tBu), 1.41 (d, $J=6.3$ Hz, 3H, CH(CH₃)OTBS) 0.90 (s, 9H, SiMe₂tBu), 0.06 (s, 3H, SiMe₂tBu), -0.07 ppm (s, 3H, SiMe₂tBu); ¹³C NMR(CDCl₃): δ = 163.72, 152.94, 152.00, 147.83, 128.89, 125.44, 125.02, 121.78, 82.21, 67.17, 58.69, 53.13, 28.05, 26.02, 25.66, 18.02, -4.88, -5.03 ppm; HRMS (FAB): m/z $[M+H]^+$ calcd for $C_{22}H_{34}NO_4Si$: 404.2257; found: 404.2278.

Adduct 87: To a solution of 56 (75% ee; 20.0 mg, 4.96×10^{-2} mmol) and H-L-Val-OFm 57 (29.3 mg, 9.92×10^{-2} mmol) in CH₂Cl₂ (0.025 mL) was added Yb(OTf)₃ (6.1 mg, 9.9×10^{-3} mmol) in H₂O (0.050 mL). The reaction mixture was stirred at room temperature for 5 days, then diluted with CHCl₃ (2 mL), and the mixture was washed with brine (2 mL \times 2). The organic layers were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (30% AcOEt/hexane) to afford 87 (16.6 mg, 48%) as a colorless foam, the diastereomer of 87 (2.4 mg, 7%) as a colorless foam, the regioisomer of 87 $(2.1 \text{ mg}, 6\%)$ as a colorless foam, and the recovered 56 (2.6 mg, 13%) as a colorless foam. 87: $R_f = 0.59$ (30% AcOEt/hexane); $\left[\alpha\right]_D^{26}$ -56.0 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{v} = 3620, 2975, 1725, 1470, 1180, 1095, 1045, 900,$ 880 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.08 (s, 1H, quinoline H-3), 7.76–7.68 (m, 2H, Fm H-4 and H-5), 7.66–7.58 (m, 2H, Fm H-3 and H-6), 7.40– 7.22 (m, 4H, Fm H-1, H-2, H-7 and H-8), 6.71 (dd, J=2.4, 10.0 Hz, 1H, quinoline H-5), 6.09 (dd, $J=1.8$, 10.0 Hz, 1H, quinoline H-6), 5.01 (q, $J=$ 6.5 Hz, 1H, CH(CH₃)OTBS), 4.80 (brs, 1H, NH), 4.66 (d, $J=12.0$ Hz, 1H, quinoline H-8), 4.53 (d, $J=6.5$ Hz, 2H, Fm CH₂), 4.21 (t, $J=6.5$, 1H, Fm H-9), 3.40 (ddd, $J=1.8$, 2.4, 12.0 Hz, 1H, quinoline H-7), 3.29 (d, $J=$ 5.6 Hz, 1 H, Val H- α), 2.43 (s, 1 H, OH), 1.95 (dqq, $J=5.6$, 6.8 Hz, 1 H, Val H- β), 1.61 (s, 9H, CO₂tBu), 1.41 (d, J = 6.5 Hz, 3H, CH(CH₃)OTBS), 0.96 (d, J=6.8 Hz, 3H, Val Me-b), 0.94 (d, J=6.8 Hz, 3H, Val Me-b), 0.91 (s, 9H, SiMe₂tBu), 0.07 (s, 3H, SiMe₂tBu), -0.05 ppm (s, 3H, Si-Me₂tBu); ¹³C NMR(CDCl₃): δ = 174.82, 163.79, 156.11, 150.50, 145.95, 143.69, 141.27, 135.02, 127.62, 127.03, 126.99, 126.08, 124.94, 121.50, 120.68, 119.87, 81.86, 72.73, 67.66, 65.98, 64.48, 59.30, 46.99, 31.85, 28.06, 25.68, 19.47, 18.25, 18.05, -4.91, -5.03 ppm; HRMS (FAB): m/z [M+ H]⁺ calcd for C₄₁H₅₅N₂O₆Si: 699.3829; found: 699.3807. Diastereomer of **87**: $R_f = 0.54$ (30% AcOEt/hexane); ¹H NMR (CDCl₃): $\delta = 8.08$ (s, 1H, quinoline H-3), 7.78–7.69 (m, 2H, Fm H-4 and H-5), 7.67–7.57 (m, 2H, Fm H-3 and H-6), 7.42–7.20 (m, 4H, Fm H-1, H-2, H-7 and H-8), 6.68 (dd, $J=2.5$, 10.4 Hz, 1H, quinoline H-5), 5.92 (dd, $J=1.8$, 10.4 Hz, 1H, quinoline H-6), 5.01 (q, $J=6.3$ Hz, 1H, CH(CH₃)OTBS), 4.78 (brs, 1H, NH), 4.72 (d, J=11.2 Hz, 1H, quinoline H-8), 4.55 (dd, J=6.4, 10.8 Hz, 1H, Fm CH₂), 4.49 (dd, $J=6.4$, 10.8 Hz, 1H, Fm CH₂), 4.22 (dd, $J=6.4$, 6.4 Hz, 1 H, Fm H-9), 3.66 (d, $J = 5.8$ Hz, 1 H, Val H- α), 3.55 (ddd, $J = 1.8$, 2.5, 11.2 Hz, 1H, quinoline H-7), 2.08–1.76 (m, 2H, Val H-b and OH), 1.62 (s, 9H, CO₂tBu), 1.40 (d, $J=6.3$ Hz, 3H, CH(CH₃)OTBS), 0.97 (d, $J=6.8$ Hz, 3H, Val Me- β), 0.91 (d, $J=6.8$ Hz, 3H, Val Me- β), 0.90 (s, 9H, SiMe₂tBu), 0.06 (s, 3H, SiMe₂tBu), -0.06 ppm (s, 3H, SiMe₂tBu). Regioisomer of 87: $R_f = 0.41$ (30% AcOEt/hexane); ¹H NMR (CDCl₃): δ = 8.10 (s, 1H, quinoline H-3), 7.78–7.69 (m, 2H, Fm H-4 and H-5), 7.66–7.58 (m, 2H, Fm H-3 and H-6), 7.42–7.18 (m, 4H, Fm H-1, H-2, H-7 and H-8), 6.56 (dd, $J=2.0$, 10.4 Hz, 1H, quinoline H-5), 6.24 (dd, $J=1.8$, 10.4 Hz, 1H, quinoline H-6), 5.07 (q, $J=6.4$ Hz, 1H, CH(CH₃)OTBS), 4.68 (dd, $J=6.2$, 10.8 Hz, 1H, Fm CH₂), 4.50–4.37 (m, 2H, quinoline H-7, Fm CH₂), 4.22 (dd, J = 6.2, 6.2 Hz, 1H, Fm H-9), 3.87 (d, J = 12.0 Hz, 1H, quinoline H-8), 3.69 (d, $J=4.4$ Hz, 1H, NH), 2.96 (d, $J=3.0$ Hz, 1H, Val H- α), 2.20–2.02 (m, 1H, Val H- β), 1.55 (s, 9H, CO₂tBu), 1.33 (d, J= 6.4 Hz, 3 H, CH(CH₃)OTBS), 1.01 (d, $J=6.8$ Hz, 3 H, Val Me- β), 0.92 (d, $J=6.8$ Hz, 3H, Val Me- β), 0.91 (s, 9H, SiMe₂t_{Bu}), 0.06 (s, 3H, Si-Me₂tBu), -0.04 ppm (s, 3H, SiMe₂tBu). H-L-Val-OFm 57 was prepared as follows: To a solution of L-valine $(3.29 \text{ g}, 28.1 \text{ mmol})$ and 1m aqueous NaOH (28.0 mL) in 1,4-dioxane (60 mL) and $H₂O$ (30 mL) was added Boc₂O (7.10 mL, 30.9 mmol). The reaction mixture was stirred at room temperature for 3 h and washed with $Et₂O$ (50 mL \times 3). The aqueous layer was acidified with 1_M aqueous HCl (30 mL) at 0° C and extracted with Et₂O (50 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated to afford Boc-l-Val-OH (6.10 g, quantitative yield) as a colorless syrup. To a solution of Boc-l-Val-OH $(1.67 \text{ g}, 7.69 \text{ mmol})$ and 9-fluorenylmethanol $(1.51 \text{ g}, 7.69 \text{ mmol})$ in CH_2Cl_2 (30 mL) were added DMAP (93.9 mg, 7.69×10^{-1} mmol) and DCC (1.59 g, 7.69 mmol). After stirring at room temperature for 2 h, the reaction mixture was evaporated and AcOEt was added to the residue. The suspension was filtered through celite and the filirate was evaporated. The residue was chromatographed on silica gel (8% AcOEt/hexane) to afford Boc-l-Val-OFm (2.76 g, 91%) as colorless solids. To a solution of Boc-L-Val-OFm (39.3 mg, 9.92×10^{-2} mmol) in CH₂Cl₂ (0.5 mL) at 0^oC was added TFA (0.5 mL). The reaction mixture was stirred at room temperature for 1 h and evaporated. The residue was dissolved in $Et₂O$ (3 mL) and this was basified with aqueous NaHCO₃, and the mixture was extracted with Et₂O (3 mL \times 3). The combined extracts were dried over Na2SO4, filtered through celite, and evaporated to afford H-l-Val-OFm **57** (29.3 mg, quantitative yield) as a colorless syrup: ${}^{1}H NMR$ (CDCl₃): δ = 7.80–7.72 (m, 2H, Fm H-4 and H-5), 7.65–7.54 (m, 2H, Fm H-3 and H-6), 7.46–7.24 (m, 4H, Fm H-1, H-2, H-7 and H-8), 4.58–4.42 (m, 2H, Fm CH₂), 4.21 (br dd, $J=6.2$, 6.2 Hz, 1H, Fm H-9), 3.31 (d, $J=5.0$ Hz, 1H, H-α), 1.97 (m, 1H, H-β), 0.92 (d, $J=6.8$ Hz, 3H, Me-β), 0.81 ppm $(d, J=6.8 \text{ Hz}, 3H, \text{ Me-}\beta).$

Segment C (6) : To a solution of 87 (27.4 mg, 3.92×10^{-2} mmol) in dry CH₂Cl₂ (0.39 mL) at 0 °C under Ar atmosphere were added 2,6-lutidine $(0.0457 \text{ mL}, \quad 7.84 \times 10^{-2} \text{ mmol})$ and TBSOTf $(0.0270 \text{ mL}, \quad 4.70 \times$ 10^{-2} mmol). After stirring at 0° C for 15 min, the reaction mixture was quenched with H₂O (2 mL) and saturated aqueous NaHCO₃ (0.50 mL). The mixture was extracted with CHCl₃ (3 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (10% AcOEt/hexane) to afford the TBS ether (30.5 mg, 96%) as a colorless syrup. To a solution of TBS ether (20.6 mg, 2.53×10^{-2} mmol) in dry CH₂Cl₂ (0.25 mL) at 0 °C under Ar atmosphere was added B-bromocatecholborane (10.1 mg, $5.07 \times$ 10^{-2} mmol). After stirring at room temperature for 1 day, the reaction mixture was quenched with $H₂O$ (2 mL). The mixture was extracted with CHCl₃ (3 mL \times 1) and AcOEt (3 mL \times 2). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (3% MeOH/CHCl₃) to afford carboxylic acid 6 (15.1 mg, 79%) as a colorless syrup: $R_f = 0.60$ (10% MeOH/ CHCl₃); $[\alpha]_D^{27}$ -113.7 (c 1.00, CHCl₃); IR (KBr): $\tilde{\nu} = 2955$, 2925, 2360, 2855, 1775, 1725, 1260, 1140, 1100, 840, 740 cm⁻¹; ¹H NMR (CDCl₃): δ = 8.28 (s, 1H, quinoline H-3), 7.81–7.68 (m, 2H, Fm H-4 and H-5), 7.64– 7.52 (m, 2H, Fm H-3 and H-6), 7.45–7.22 (m, 4H, Fm H-1, H-2, H-7 and H-8), 6.76 (dd, $J=0.8$, 10.2 Hz, 1H, quinoline H-5), 6.15 (dd, $J=3.4$, 10.2 Hz, 1H, quinoline H-6), 5.09 (q, $J=6.1$ Hz, 1H, CH(CH₃)OTBS), 4.69 (d, $J=7.6$ Hz, 1H, quinoline H-8), 4.56 (dd, $J=6.3$, 10.8 Hz, 1H, Fm CH₂), 4.50 (dd, $J=6.3$, 10.8 Hz, 1H, Fm CH₂), 4.17 (dd, $J=6.3$, 6.3 Hz, 1H, Fm H-9), 3.47 (ddd, J=0.8, 3.4, 7.6 Hz, 1H, quinoline H-7), 3.10 (d, $J=5.5$ Hz, 1H, Val H- α), 1.81 (dqq, $J=5.5$, 7.0, 7.0 Hz, 1H, Val H- β), 1.37 (d, $J=6.1$ Hz, 3H, CH(CH₃)OTBS), 0.93 (s, 9H, SiMe_{2t}Bu), 0.89 (s, 9H, SiMe₂t_{Bu}), 0.85 (d, J = 7.0 Hz, 3H, Val Me- β), 0.79 (d, J = 7.0 Hz, 3H, Val Me- β), 0.16 (s, 3H, Si Me_2tBu), 0.06 (s, 3H, Si Me_2tBu), 0.04 (s, 3H, Si \underline{Me}_2tBu), -0.04 ppm (s, 3H, Si \underline{Me}_2tBu); ¹³C NMR(CDCl₃): $\delta=$ 174.55, 164.18, 155.82, 152.38, 143.55, 143.22, 141.36, 133.38, 128.25, 127.81, 127.10, 127.06, 124.84, 124.75, 121.33, 120.50, 120.03, 74.04, 67.29, 66.02, 63.80, 56.95, 47.01, 31.73, 25.74, 25.69, 25.58, 19.37, 18.16, 4.23, -4.53, -4.87, -4.95 ppm; HRMS (FAB): m/z $[M+H]^+$ calcd for $C_{43}H_{61}N_2O_6Si_2$: 757.4068; found: 757.4076.

Phenylselenoalanine 89: To a solution of 88 (1.00 g, 5.34 mmol) in degassed dry DMF (13 mL) at room temperature under Ar atmosphere was added benzeneselenol (0.681 mL, 6.41 mmol). After stirring at room temperature for 2 h, the reaction mixture was quenched with 1m aqueous NaOH (6 mL) and H₂O (10 mL), and the mixture was washed with $Et₂O$ $(10 \text{ mL} \times 3)$. The aqueous layer was acidified with 1m aqueous HCl to pH 2 at 0° C and extracted with AcOEt (30 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel $(0-5\% \text{ MeOH/CHCl}_3)$ to afford 89 (1.84 g, quantitative yield): $R_f = 0.61$ (10% MeOH/CHCl₃); ¹H NMR (CDCl₃): δ = 7.60–7.48 (m, 2H, Ph), 7.30–7.18 (m, 3H, Ph), 5.50 (br d, $J=7.2$ Hz, 1 H, BocNH), 4.70–4.56 (m, 1 H, H- α), 3.44 (dd, $J=12.0$, 4.6 Hz, 1 H, H- β), (dd, $J=12.0$, 5.2 Hz, 1 H, H- β), 1.40 ppm (s, 9 H, Boc). Phenylselenoalanine Fm ester 90: To a solution of 89 (996 mg, 2.89 mmol) and 9-fluorenylmethanol (568 mg, 2.89 mmol) in dry CH_2Cl_2 (15 mL) at 0° C under Ar atmosphere were added DMAP (35.3 mg, 2.89×10^{-1} mmol) and DCC (597 mg, 2.89 mmol). After stirring at room temperature for 4 h, the reaction mixture was evaporated and AcOEt was added to the residue. The suspension was filtered through celite and the filtrate was evaporated. The residue was chromatographed on silica gel (10% AcOEt/hexane) to afford 90 (1.23 g, 82% from 88): $R_f = 0.61$ (20% AcOEt/hexane); m.p. 109-110 °C (not recrystallized); $\left[\alpha\right]_D^{29}$ + 18.6

 $(c 1.00, CHCl₃)$; IR $(CHCl₃)$: $\tilde{v} = 3620, 3435, 2980, 1765, 1710, 1580, 1420,$ 1300, 1130, 875 cm⁻¹; ¹H NMR (CDCl₃): δ = 7.80–7.72 (m, 2H, Fm), 7.57– 7.15 (m, 11H, Fm and PhSe), 5.34 (br d, J=8.0 Hz, 1H, NHBoc), 4.72 (dt, $J=5.0$, 8.0 Hz, 1H, H- α), 4.24 (br dd, $J=6.5$, 9.2 Hz, 1H, Fm C \underline{H}_2), 4.14–4.01 (m, 2H, Fm CH₂ and H-9), 3.28 (d, $J=5.0$ Hz, 2H, H- β), 1.42 ppm (s, 9H, Boc); 13 C NMR (CDCl₃): δ = 170.57, 154.92, 143.36, 143.24, 141.24, 141.17, 133.67, 129.10, 128.78, 127.83, 127.50, 127.12, 127.09, 125.03, 124.93, 120.00, 119.97, 80.06, 67.16, 53.28, 46.49, 30.50, 28.23 ppm; HRMS (FAB): m/z $[M+H]^+$ calcd for $C_{28}H_{30}NO_4^{80}Se$: 524.1340; found: 524.1342.

Dipeptide 91: To a solution of 90 (1.23 g, 2.35 mmol) in dry CH_2Cl_2 (6.0 mL) at 0° C under Ar atmosphere was slowly added TFA (6.0 mL) . The reaction mixture was stirred at room temperature for 2 h and evaporated to afford the crude amine·TFA. To a solution of this crude amine·T-FA in dry CH_2Cl_2 (12 mL) at 0°C under an Ar atmosphere were added iPr2NEt (1.00 mL, 5.74 mmol), 89 (895 mg, 2.60 mmol), HOAt (354 mg, 2.60 mmol), and CIP (724 mg, 2.60 mmol). After stirring at room temperature for 2 h, the reaction mixture was quenched with H_2O (10 mL) and saturated aqueous NaHCO₃ (2 mL) . The mixture was extracted with CHCl₃ (15 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated. The residue was chromatographed on silica gel (20% AcOEt/hexane) to afford 91 (1.66 g, 94%) as a colorless foam: $R_f=0.59$ (40% AcOEt/hexane); m.p. 120 °C (not recrystallized); $[\alpha]_D^{31}$ 0.00 (c 1.00, CHCl₃); IR (CHCl₃): $\tilde{\nu} = 3620, 3420, 2975, 1765,$ 1710, 1680, 1580, 1475, 1420, 1370, 875 cm⁻¹; ¹H NMR (CDCl₃): δ = 7.79– 7.15 (m, 18H, Fm and PhSe \times 2), 6.95 (br d, J = 7.0 Hz, 1H, CONH), 5.05 (brs, 1H, NHBoc), 4.82 (ddd, $J=4.6$, 4.6, 7.0 Hz, 1H, PhSeAla H- α), 4.34–4.16 (m, 1H, PhSeAla H- α), 4.27 (dd, J=6.6, 10.0 Hz, 1H, Fm CH₂), 4.10 (dd, $J=6.6$, 10.0 Hz, 1H, Fm CH₂), 4.02 (dd, $J=6.6$, 6.6 Hz, 1H, Fm H-9), 3.31 (dd, $J=4.6$, 12.4 Hz, 1H, PhSeAla H- β), 3.24–3.04 (m, 3H, PhSeAla H- $\beta \times 3$), 1.43 ppm (s, 9H, Boc); ¹³C NMR (CDCl₃): δ = 169.96, 169.66, 155.05, 143.26, 143.13, 141.24, 141.15, 133.70, 133.04, 129.15, 128.78, 127.86, 127.60, 127.37, 127.16, 127.12, 124.93, 124.85, 120.00, 80.39, 67.28, 54.18, 52.46, 46.40, 29.69, 29.56, 28.23 ppm; HRMS (FAB): m/z [M+H]⁺ calcd for C₃₇H₃₉N₂O₅⁸⁰Se₂: 751.1189; found: 751.1174.

Segment D (7): Compound 91 (480 mg, 6.41×10^{-1} mmol) in 3M HCl/ AcOEt (2.9 mL) was stirred at room temperature for 2 h and the solvent was evaporated. The residue was dissolved in AcOEt (2 mL) and this was basified with aqueous NaHCO₃, and the mixture was extracted with AcOEt (3 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated to afford 7 (416 mg, quantitative yield), which was used for the next step without purification.

Dipeptide 92: To a solution of 89 (1.05 g, 3.05 mmol) in dry THF (30 mL) at -15° C under Ar atmosphere were added NEt₃ (0.447 mL, 3.21 mmol) and ClCO₂Et (0.306 mL, 3.20 mmol). After stirring at -15° C for 10 min, 28% aqueous NH₃ (6.2 mL, 90 mmol) was slowly added. The reaction mixture was stirred for 30 min at -15° C and then at room temperature for 2 h. The reaction mixture was evaporated and H_2O (20 mL) was added. The mixture was extracted with AcOEt $(30 \text{ mL} \times 3)$. The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residue was chromatographed on silica gel (50% AcOEt/hexane) to afford the amide (826 mg, 79%) as a colorless syrup: R_f =0.18 (50% AcOEt/hexane); ¹H NMR (CDCl₃): δ =7.59–7.53 (m, 2H, PhSe), 7.32-7.22 (m, 3H, PhSe), 6.40 (brs, 1H, CONH₂), 5.98 (brs, 1H, CONH₂), 5.39 (br d, $J=6.8$ Hz, 1H, BocNH), 4.37 (m, 1H, H- α), 3.25 (m, 2H, H- $\beta \times 2$), 1.42 ppm (s, 9H, Boc). To a solution of this amide (75.3 mg, 2.08×10^{-1} mmol) in dry CH₂Cl₂ (2.0 mL) at 0°C under Ar atmosphere was slowly added TFA (0.5 mL). The reaction mixture was stirred at room temperature for 1 h and evaporated to afford the crude amine·TFA (102 mg). To a solution of this crude amine·TFA in dry DMF (2.2 mL) at 0°C under Ar atmosphere were added iPr_2NEt (0.151 mL, 8.67 x 10^{-1} mmol), 89 (82.3 mg, 2.40×10^{-1} mmol), HOAt (32.5 mg, 2.39 \times 10^{-1} mmol), and CIP (66.6 mg, 2.39×10^{-1} mmol). After stirring at room temperature for 5 h, the reaction mixture was quenched with H_2O (4 mL). The mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over $Na₂SO₄$, filtered through celite, and evaporated. The residual solid was recrystallized from acetone/hexane to

afford 92 (115 mg, 92%) as colorless crystals: $R_f = 0.78$ (10% MeOH/ CHCl₃); m.p. 167–168 °C; $\left[\alpha\right]_D^{23}$ –72.9 (c 1.02, CHCl₃); IR (KBr): \tilde{v} = 3320, 3195, 2980, 1690, 1655, 1625, 1525, 1480, 1385, 1165, 735, 690, 670 cm⁻¹; ¹H NMR (CDCl₃): δ = 7.59–7.43 (m, 4H, PhSe), 7.33–7.21 (m, 6H, PhSe), 6.93 (brd, $J=8.4$ Hz, 1H, CONH), 6.72 (brs, 1H, CONH₂), 5.44 (brs, 1H, CONH₂), 5.05 ppm (brd, $J=5.4$ Hz, 1H, NHBoc); ¹³C NMR $(CDCl_3)$: $\delta = 172.04$, 170.22, 155.80, 133.31, 133.07, 129.43, 129.28, 127.91, 127.63, 81.21, 54.79, 52.74, 29.10, 28.74, 28.25 ppm; HRMS (ESI): m/z $[M+Na]^+$ calcd for $C_{23}H_{29}N_3NaO_4^{80}Se_2$: 594.0386; found: 594.0388.

Segment E (4) : To a solution of 92 (47.1 mg, 8.27×10^{-2} mmol) in dry CH₂Cl₂ (0.67 mL) at 0 °C under Ar atmosphere was slowly added TFA (0.67 mL). The reaction mixture was stirred at room temperature for 2 h and evaporated. The residue was dissolved in AcOEt (5 mL) and this was basified with aqueous NaHCO₃, and the mixture was extracted with AcOEt (5 mL \times 3). The combined extracts were dried over Na₂SO₄, filtered through celite, and evaporated to afford 4 (36.1 mg, 93%) as colorless solids, which was used for the next step without purification.

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